

ECOPHYSIOLOGICAL RESPONSES OF TWO TROPICAL URBAN SHRUB PLANTS TO ELEVATED CARBON DIOXIDE AND WATER STRESS

WAN NUR AIN, N. and WAN JULIANA, W.A.*

School of Environmental and Natural Resource Sciences, Faculty of Science and Technology,
Universiti Kebangsaan Malaysia, 43600 Bangi, Selangor, Malaysia

*E-mail: ayie@ukm.edu.my

Accepted 1 February 2017, Published online 31 March 2017

ABSTRACT

Elevated atmospheric carbon dioxide causes changes in most plant species in terms of physiological and morphological properties. In this study, we elucidate the effects of elevated CO₂ and water regimes on two species of shrubs which were *Melastoma malabathricum* and *Clidemia hirta*. The factor of water stress was also included whereby for each CO₂ treatment, the amount of water was given once or twice daily. The treatment of elevated CO₂ was at 800 ppm, where the plants were exposed for two hours daily and ambient CO₂ at 400 ppm as the control. The growth of plants was monitored through their height, leaf area and biomass that were recorded fortnightly for six months. The physiological changes were determined via stomatal conductance, photosynthetic rate and transpiration rate. The results revealed that each species responded differently to the CO₂ treatments. The growth parameters of height and leaf area index showed positive responses on elevated CO₂ and twice watering for both species. As for biomass, *M. malabathricum* exhibited higher below ground biomass compared to above ground biomass in the elevated CO₂. In contrast, *C. hirta* showed the opposite trend. The results of the three physiological parameters demonstrated a bell shape trend for both treatments with the highest rate at the fifth month.

Key words: elevated CO₂, shrubs, photosynthesis, growth, tropical plant

INTRODUCTION

Climate change is principally driven by the burning of fossil fuels and greenhouse gas emissions from deforestation, degradation and land use change. As a result of the latter, the earth's natural capital is continuing to be badly degraded and thus affected water availability, soil stability, air quality, and forests ecosystem balance (International Sustainability Unit, 2015). The impacts of climate change has been considered as national security issue and may have many other indirect negative effects for livelihood and economy (Barnett, 2001). Enormous research efforts have been undertaken to understand how plants and ecosystems, both natural and managed, respond to rising CO₂ (Ainsworth & Long, 2005) and water stress (Corlett, 2016). For example, primary effects of rising CO₂ have been well documented and include reduction in stomatal conductance and transpiration, improved water-use efficiency, higher rates of photosynthesis and

increased light-use efficiency of plants (Drake *et al.*, 1997). Whereas, plants under water stress were reported contribute an imbalance in biomass partition and also the productivity of the plants (Bastin *et al.*, 2015; Fauset *et al.*, 2015).

Effects of climate change and increased of carbon dioxide on the growth of individual plant species have been conducted on various kinds of plants. Most of the studies focused on crop plants such as *Ribes nigrum* and *Hordeum vulgare* L. (barley) (Johnson *et al.*, 2011; Robredo & Usue, 2007; Rogers *et al.*, 1996; Shimono *et al.*, 2013; Wang *et al.*, 2015), horticultural plants (Xu *et al.*, 2014), fruit trees (Schaffer *et al.*, 1999) and understory herbs native to Europe (Anderson & Cipollini, 2013). Most plants reacted positively to the increase in carbon dioxide (Niell, 2003) and also to other factors such as the content of nitrogen (Geisinger *et al.*, 1998), water regime, temperature and others. However, the effects of elevated CO₂ may vary across season (Anderson & Cipollini, 2013) and species itself. Rogers *et al.* (1996) reported that, exposure of plant canopies to high concentration of

* To whom correspondence should be addressed.

carbon dioxide often stimulates the growth of both shoots and roots of the plants but there may be differences in allocation between root and shoot, at least for certain conditions. However, little did we know about the study involving tropical shrubs.

Shrubs are important for moderating a range of ecosystem services because they influence micro-climate, reduce erosion and nutrient loss, increase water holding capacity, maintain soil structure and stability and provide habitat for a range of taxa (Maestre *et al.*, 2009). It has been proposed that disturbances ranging from climate change to grazing to fire suppression can generate heterogeneity in soil resources and thus create opportunities for shrub colonization (Schlesinger *et al.*, 1990).

Melastoma malabathricum L. and *Clidemia hirta* L. from *Melastomataceae* family are amongst the widely distributed shrubs in tropical open areas or urban forests. *M. malabathricum* can be found abundantly in Malaysia. Additionally, this plant has been reported to be found growing wild in the Indian Ocean Islands, throughout South and South-East Asia, China, Taiwan, Australia and the South Pacific Ocean (Wong, 2008). This species is a common herbal plant used in folk treatment to treat inflamed wounds (Omar, 2012), while leaf, root and also flower crude from this species had been used to treat toothache, diarrhea, scar prevention, wounds, post-partum recovery and others (Joffry *et al.*, 2012). As for *C. hirta*, its natural range is extremely wide, extending throughout most of the humid tropical portions of Central and South America from Southern Mexico to Northern Argentina and the islands of the West Indies. It is characteristically found in moist, shaded localities on the edges of clearings and stream-banks, in ditches, and along paths and roadways (Wester & Wood, 1977). Medicinally, *C. hirta* is the second plant preferred after *Anacardium occidentale* L. to treat leishmanial ulcers (Franca *et al.*, 1996).

In this study, we examined the effects of elevated CO₂ and water regime on two tropical shrub species of *M. malabathricum* and *C. hirta* in terms of growth and physiology. The two shrub species were selected because they grow wild and commonly found in urban forest areas. Since both selected plant species are widely dispersed at urban forested areas, we hypothesized that elevated carbon dioxide concentration in their surroundings will positively change the plant growth morphologically and physiologically and will in turn affect their abundance in the urban forest areas. Meanwhile, the different water regime is expected to reveal the physiological mechanisms of the two tropical shrub plants adapting to water stress.

MATERIALS AND METHODS

Plant materials

Seedlings of *M. malabathricum* were obtained from seeds that were collected from mature plants which grew wild along the roadsides of UKM Bangi campus (2.9192° N, 101.7691° E). Meanwhile, for *C. hirta*, stem cuttings of the mature plants that grew at similar location were used instead, after several attempts made to obtain seedlings from its seeds were unsuccessful. The seeds and the stem cuttings were then planted in nursery trays filled with topsoil and watered daily until the seedlings emerged which, were around seven to ten days after planted. The growing medium used was topsoil of podzolic type mixed with organic fertilizer with a ratio of 4:1. After the saplings reached the age of 2 weeks old, each sapling were planted in a 8 x 6 inches polybag with NPK fertilizer at the ratio of 15:15:15 at a low dose of 5 g/polybag/month.

Experimental design

The plants were then divided into four groups of treatments. Two factors taken into account were the concentration of carbon dioxide and water regime. Each factor had 10 replications of saplings. The concentration of carbon dioxide for ambient as the control was at 400±50 ppm and elevated CO₂ was at 800±50 ppm. For the ambient CO₂ treatment, the saplings were grown in a shade house. Saplings exposed to elevated CO₂ were in an open roof greenhouse. The elevated CO₂ was injected for two hours from 0900 am until 1100 am, daily. The roof is covered with a double layer of UV protected polycarbonate material. The side wall is covered with mesh netting and insect-proofed. A centralized control panel is installed to control all of the systems and monitor relative humidity, temperature and carbon dioxide concentration in the greenhouse. For each treatment of carbon dioxide, saplings were divided into two water regimes where they were either watered once or twice daily. The amount of water given each time was approximately 500 mL.

Growth measurements

Growth parameters of each sapling were measured fortnightly for six months from April 2015 until September 2015. The growth parameters presented here are height, leaf area index and biomass as these growth measurements reflected the percentage of growth for the plants in the experiment by analyzing the trends of its increment. However, the three parameters are inter-related. For example, the increment in height and leaf area will thus affect its biomass composition. As for the leaf area, all parts

of the leaves were laid on a paper then digital images were taken with a ruler as the reference scale. The leaf area was next calculated using the software of ImageJ. Measurements for stem height were made from the base of the stem up to its apex. At the end of the treatment, all of the plants were harvested and both the above ground and below ground biomass were obtained. The fresh weight of biomass was measured using a digital scale up to two decimal points. Dry weight measurements were conducted after the plants were dried in the oven at a temperature of 65°C for seven to ten days.

Physiological parameters

Physiological changes of plants were monitored using the photosynthetic rate, transpiration rate and stomatal conductance. These parameters were chosen because photosynthetic rate do affects the growth of the plant (Kirschbaum, 2011) meanwhile, stomatal conductance is closely related to the rate of photosynthesis (Farquhar & Sharkey, 1982) so as the transpiration rate. For this physiological data, we used portable infrared gas analyser (Li-6400, Li-Cor, Lincoln, NE, USA).

Statistical analysis

Differences between means of all measured parameters were tested using the General Linear Model (GLM) in Minitab version 17 at the significance level of 5% ($p < 0.05$).

RESULTS AND DISCUSSION

Effect of Elevated CO₂ and Water Regime on Plant Height

Plants of *M. malabathricum* in ambient and elevated carbon dioxide showed a significant different ($p < 0.05$) where the height of *M. malabathricum* was higher at the elevated than the one in ambient CO₂ (Figure 1). However, the water treatment did not show a significant different of plant heights ($p = 0.34$). The stem height of *M. malabathricum* showed the highest increment for plants grown in elevated CO₂ with 240% and 193% of height increment for once and twice watered, respectively. Meanwhile, in the ambient CO₂ the height increment was 106% and 107% for once and twice watered, respectively. Our results clearly indicated that *M. malabathricum* stem growth reacted positively with the increase of CO₂ concentration.

Contrary to the *M. malabathricum*, *C. hirta* on the other hand, showed the highest increase in percentage of mean stem height were for plants grown in ambient and once watered which was 232% of height increment followed by elevated CO₂ and twice watered with percentage of increase at 174%. Meanwhile, the treatment of elevated CO₂ and once watered showed only 146% of *C. hirta* stem height increment. In short, the two selected shrub plant responded differently to the elevated

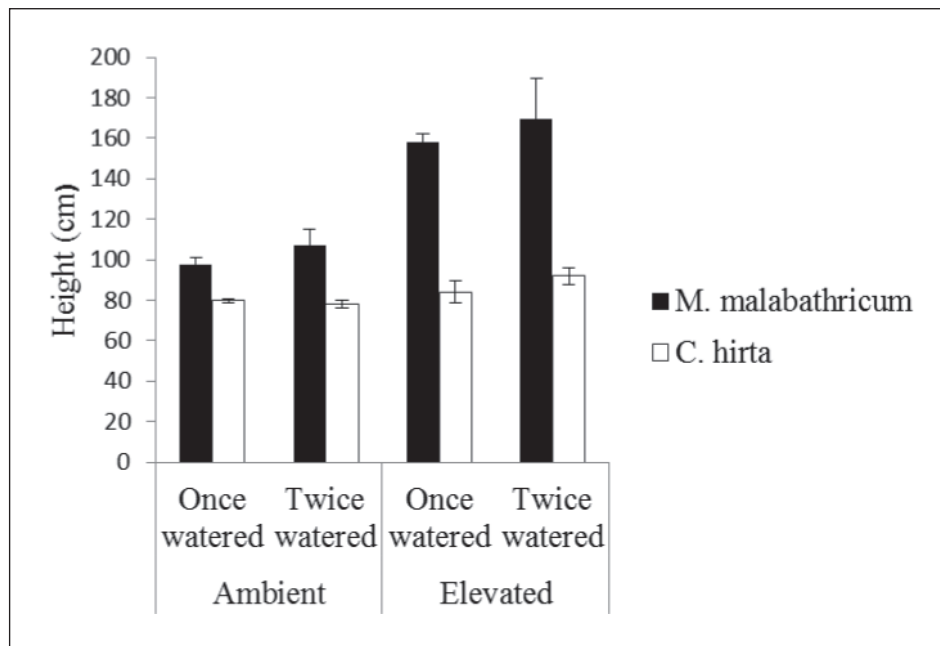


Fig. 1. Effect of elevated CO₂ concentration and water regime on the mean (\pm SE) height of *M. malabathricum* and *C. hirta* ($p < 0.05$).

CO₂. Individual plant organs have often been observed to enlarge proportionally with increasing CO₂ such as increasing stem and roots partitioning (Allen *et al.*, 1990) and also the expanse and thickness of leaves increased (Thomas & Harvey, 1983).

For the six months of the elevated CO₂ treatment, an analysis on the plant height relative growth rates (RGR) of *M. malabathricum* exhibited that at elevated CO₂, an increase of RGR was evident at the earlier treatment but after the second month, a continuous decrease of RGR was observed (Figure 2). The *M. malabathricum* plants in ambient

CO₂ also displayed similar trend except that the RGR was higher at the elevated CO₂. Although the elevated CO₂ treatment increased the RGR of *M. malabathricum* stem height and consequently the mean RGR, past studies showed that the enhancement might be reduced if the treatment is prolonged (Ceulemans & Mousseau, 1994). Meanwhile, *C. hirta* showed a high RGR of stem height for the first month but, sharp decreases of RGR for the remaining months. Compared to *M. malabathricum*, the *C. hirta* displayed insignificant differences ($p > 0.05$) between treatments of water regime and CO₂ concentration.

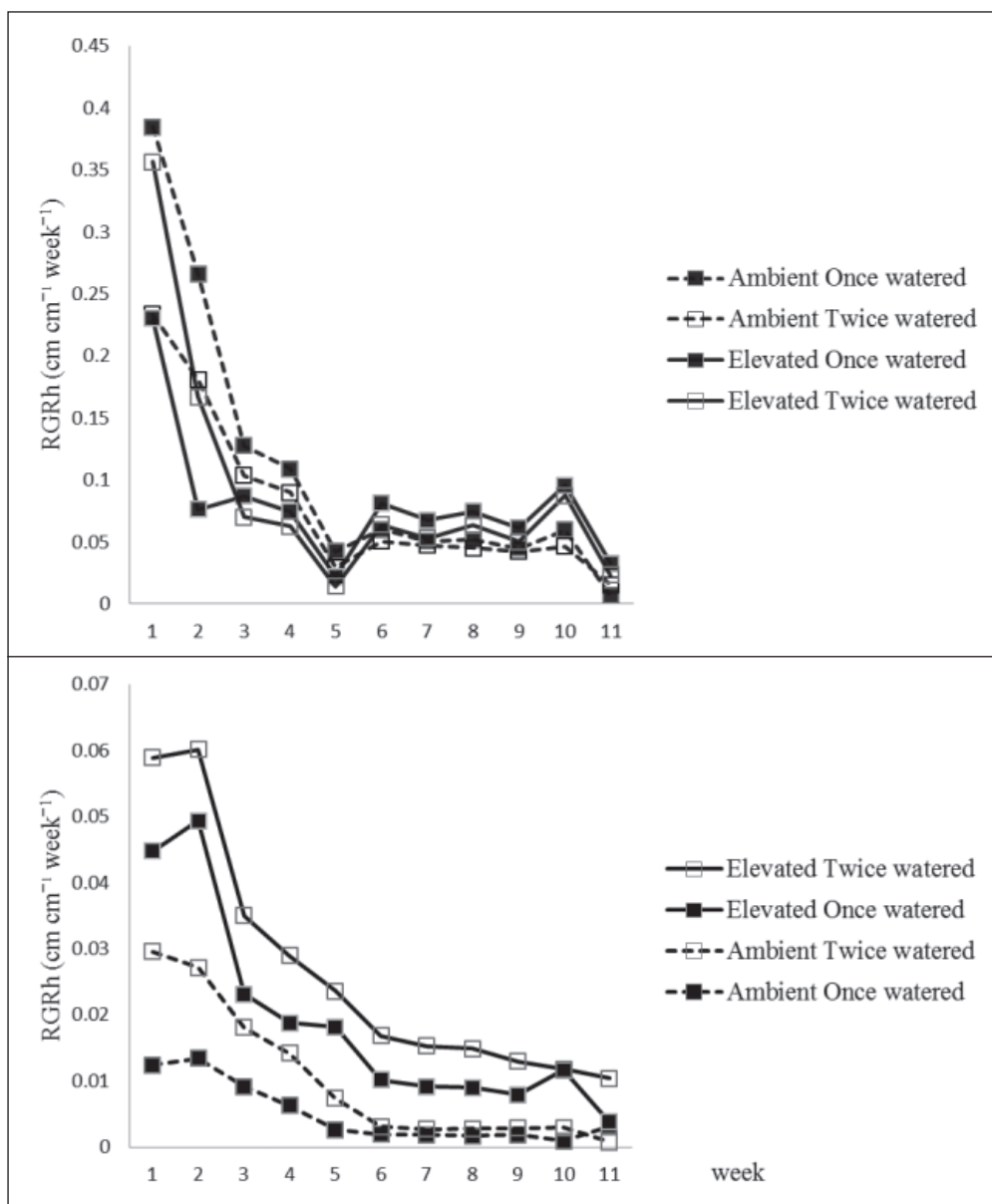


Fig. 2. Time course of relative growth rate for the height of *Clidemia hirta* (above) and *Melastoma malabathricum* (below) seedlings for eleven weeks at ambient (400±50 ppm) and elevated CO₂ (800±50 ppm) and two water regime.

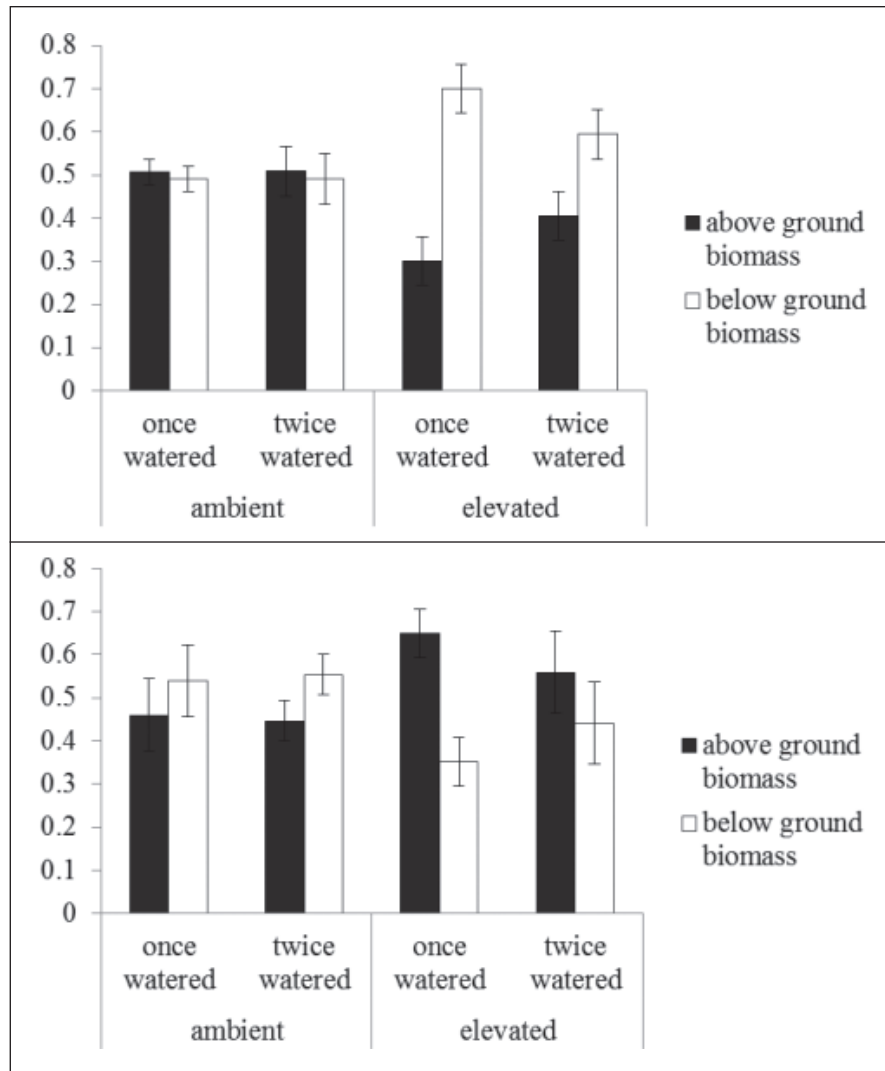


Fig. 3. Effect of elevated CO₂ concentration and water regime on the mean (\pm SE) biomass partitioning of *M. malabathricum* (above) and *C. hirta* (below) ($p < 0.05$).

Effect of Elevated CO₂ and Water Regime on Biomass Allocation

The *M. malabathricum* responded to elevated CO₂ by increasing below ground biomass. On the contrary, *C. hirta* had higher aboveground biomass in elevated rather than in ambient CO₂ (Figure 3). The differences between plant species may be due to the partitioning of photosynthetic which may be influenced by environmental stimuli as stated by Rogers *et al* (1996) and it differs for different species. Our results agrees with Farrar and Gunn (1996) that there are no reason to assume that shoots will increase or decrease more than roots. In addition, Geisinger *et al* (1995) demonstrated that root volumes for below ground growth largely mirrored that of the shoot, or at least do not deviate sufficiently to substantially modify the growth pattern. This concludes that different species offers different results in its biomass allocation.

Water regime did not significantly affect plant biomass allocation at both CO₂ concentrations for both species ($p > 0.05$). However, plants in the elevated CO₂ treatment had higher differences when compared between once and twice watered whereas, in ambient CO₂ not much differences between plants of once and twice watered. This may be due to the interaction of water regime and CO₂ treatment ($p < 0.05$). Plants exposed to higher CO₂ may have a better increase in biomass as the amount of water given was sufficient for plant growth.

Effect of Elevated CO₂ and Water Regime on Leaf Area Index

Elevated CO₂ significantly increased the leaf area index of both species. An interaction of water regime and CO₂ gave significant ($p < 0.05$) effect on the leaf area index of both species except for twice watered *C. hirta*. For *M. malabathricum*, the

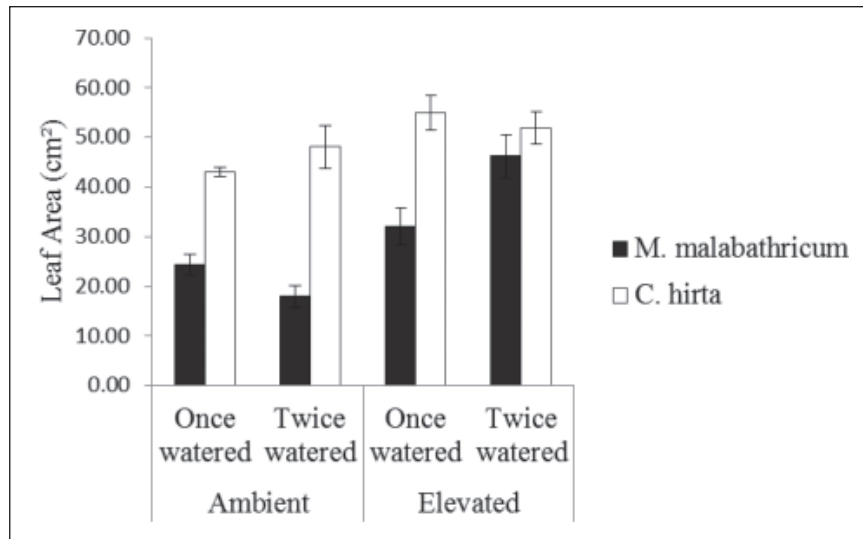


Fig. 4. Effect of elevated CO₂ concentration and water regime on the mean (\pm SE) leaf area index of *M. malabathricum* and *C. hirta* ($p < 0.05$).

highest mean leaf area was 46.24 cm² in the elevated CO₂ and twice watered followed by elevated CO₂ and once watered of 32.06 cm². Plants of *M. malabathricum* in ambient CO₂ had smaller size of leaf area with the mean value of 24.35 cm² and 17.94 cm² for once and twice watered, respectively. Our results suggested that the effect of water stress on *M. malabathricum* leaf area was more pronounced when grown in elevated than ambient CO₂.

Similar effect of interaction between CO₂ and water regime were shown by the plants of *C. hirta* except that its leaf area was bigger compared to the *M. malabathricum* (Figure 4). *C. hirta* plants in the elevated CO₂ and once watered exhibited the highest mean leaf area with the value of 55.0 cm² followed by plants in elevated CO₂ and twice watered of 51.8 cm² ($p < 0.05$). Contrary to the elevated CO₂, the *C. hirta* plants had larger size in ambient CO₂ and twice watered compared to plants in ambient and once watered with the value of 48.2 cm² and 43.00 cm², respectively ($p < 0.05$), indicating the effect of water stress on *C. hirta* leaf area was more pronounced in ambient CO₂.

Our results is in line with Sasek and Strain (1989) which revealed that changes in plant water status have been identified as mechanism for increased leaf area in elevated CO₂. Previous study on meta-analysis of soybean (*Glycine max*) showed that when this plants was exposed to elevated CO₂, their average leaf area index (LAI) increased by 18% relative to ambient CO₂ (Ainsworth *et al.*, 2002). However, the leaf area in elevated CO₂ has been found to vary both among and within plant species itself (Taylor *et al.*, 1994; Masle, 2000).

Effect of Elevated CO₂ and Water Regime on Photosynthesis Rate (P_s)

Six month's exposure of ambient and elevated CO₂ resulted in a variation of bell curve-shaped P_s of *M. malabathricum* for both once and twice watered. A higher rate for the first two months than the remaining months in the ambient CO₂ was observed for *M. malabathricum* plants (Figure 5). The highest value of P_s was the *M. malabathricum* grown in ambient and once watered at the second month with the value of 16.3 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ followed by the P_s of the first month with the value of 16.1 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$. The highest P_s in elevated CO₂ was the one with twice watered at 13.7 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ which had the same P_s at ambient and twice watered. A lower P_s in elevated CO₂ showed that *M. malabathricum* able to adapt and take an advantage on the increasing CO₂ in urban areas.

As for *C. hirta*, when the plant was exposed to elevated CO₂, over time, similar variation of bell-shaped P_s was observed with an exception of exponential P_s in elevated CO₂ and twice watered. The highest P_s of *C. hirta* were the plants in elevated CO₂ and once watered at the value of 11.1 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ at the third month followed by P_s of *C. hirta* in ambient and once watered with the value of 10.2 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ at the fourth month.

In short, the two species' P_s react differently to elevated CO₂ ($p < 0.05$). Photosynthetic rate for *M. malabathricum* was lower in elevated than in ambient CO₂ whereas for *C. hirta* the P_s was higher in elevated than in ambient CO₂. Meanwhile, water regimes did not significantly ($p = 0.10$) affect the P_s of both species at both CO₂. Our results correspond

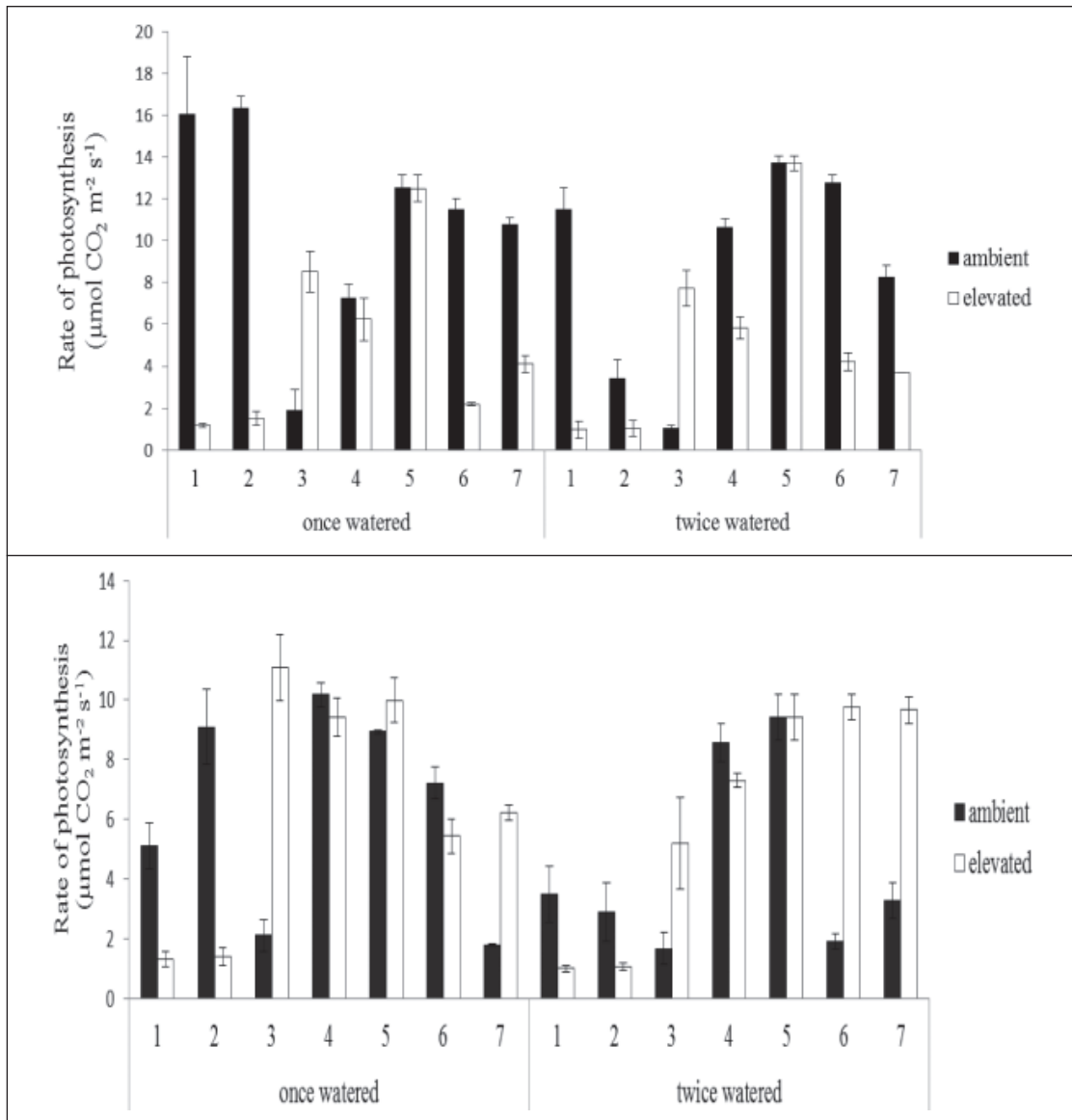


Fig. 5. Effect of elevated CO₂ concentration and water regime on the mean (\pm SE) photosynthesis rate of *Melastoma malabathricum* (above) and *Clidemia hirta* (below) ($p < 0.05$).

to a study by Nagy *et al* (2000) whereby CO₂ treatment do affects the values of P_s but not the pattern. As CO₂ increased over time, the P_s would initially increase and started to decrease after a certain period of plant growth. Our results also indicated that the interactions between species and CO₂ level do affect the rate of photosynthesis significantly rather than species-water regime and CO₂ level-water interactions.

Effect of Elevated CO₂ and Water Regime on Stomatal Conductance (g_s)

The g_s response of *M. malabathricum* during the six months CO₂ exposure was a bell curve trend for both ambient and elevated treatments. A lower rate of g_s was found for *M. malabathricum* plants grown in the elevated than ambient CO₂ ($p < 0.05$).

The highest g_s of *M. malabathricum* was on the fifth month of twice watered plants in ambient and elevated with the same value of 0.6 mol H₂O m⁻² s⁻¹. However, there was not much difference in both CO₂ treatments related to different water regime (Figure 6).

Correspondingly, for *C. hirta*, plants in elevated CO₂ also had a lower rate of g_s when compared to the one in the ambient CO₂. Our results also showed that a high g_s of *C. hirta* at the first two months in ambient CO₂ for both water regime treatments. The least g_s rate of *C. hirta* was in elevated CO₂ and twice watered. The highest g_s of *C. hirta* was the plants in ambient and twice watered with the value of 0.9 mol H₂O m⁻² s⁻¹ at the fifth month followed by the one in ambient and once watered at the second month with the value of 0.6 mol H₂O m⁻² s⁻¹.

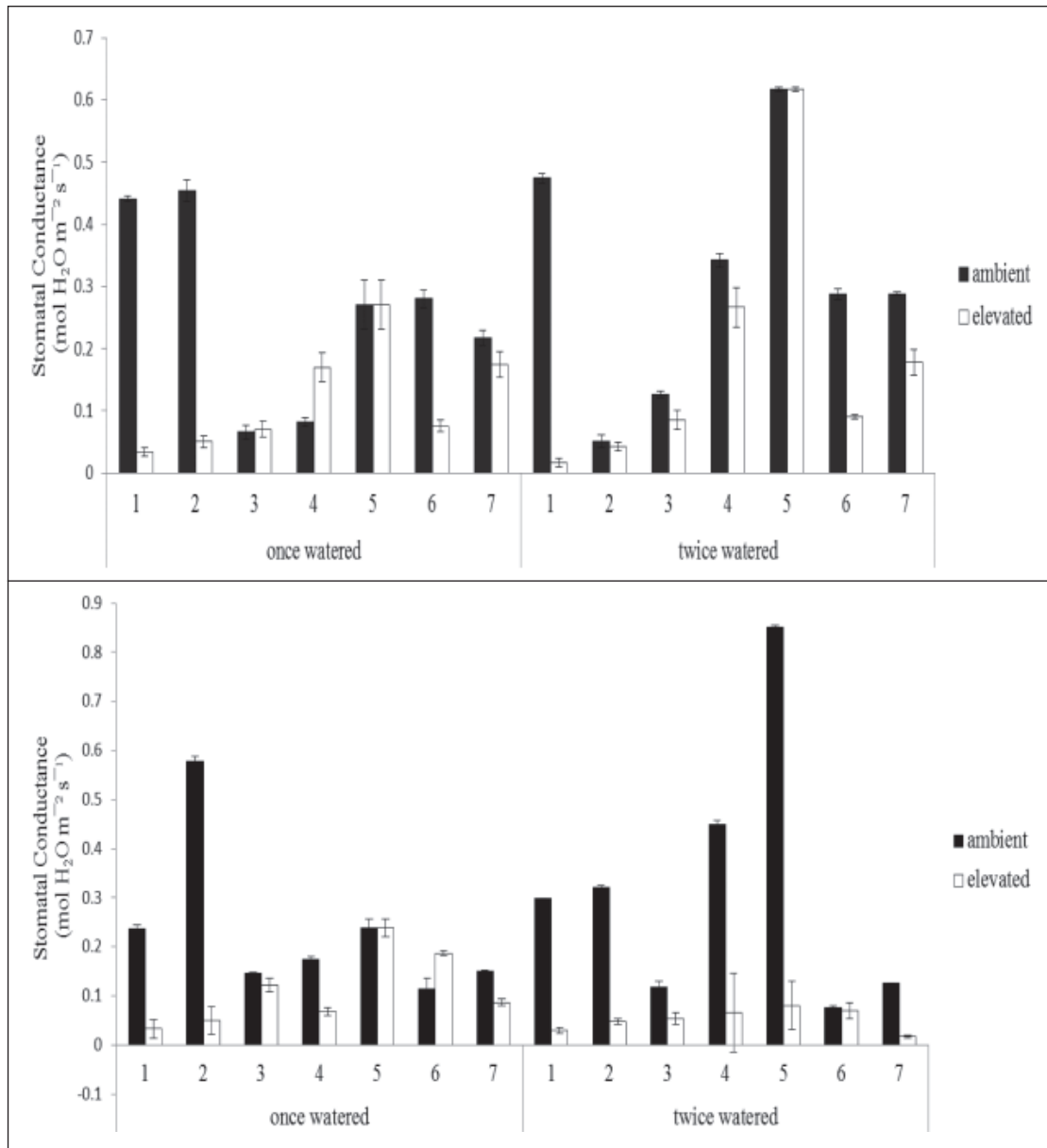


Fig. 6. Effect of elevated CO₂ concentration and water regime on the mean (\pm SE) stomatal conductance of *Melastoma malabathricum* (above) and *Clidemia hirta* (below) ($p < 0.05$).

Robredo and Usue (2007) reported that stomatal conductance decline with increasing CO₂ but the rate of reduction is higher under sufficient water rather than under water stress. The lower stomatal conductance in elevated CO₂ was also reported by Roberntz and Stockfors (1998) in their experiment on *Picea abies* (L.) Karst. They suspected that these two species are less affected by elevated CO₂ compared to the one that have higher stomatal conductance as stated by Morison (1985). A decrease in stomatal conductance in elevated CO₂ may due to a decrease in foliar nitrogen concentration in response to elevated CO₂ (Roberntz

& Stockfors 1998). Moreover, Woodrow (1994) suggested that C₃ plants possess a genetically controlled feedback mechanism to smooth out imbalances within photosynthetic system caused by a rise in atmospheric CO₂ concentration.

Effect of Elevated CO₂ and Water Regime on Transpiration Rate (E)

Responses of this two plant species transpiration rate to differences in CO₂ and water regimes showed that a lower E was observed at the elevated than ambient CO₂ ($p < 0.05$). Meanwhile, water regime did not significantly affect the E of both species at both

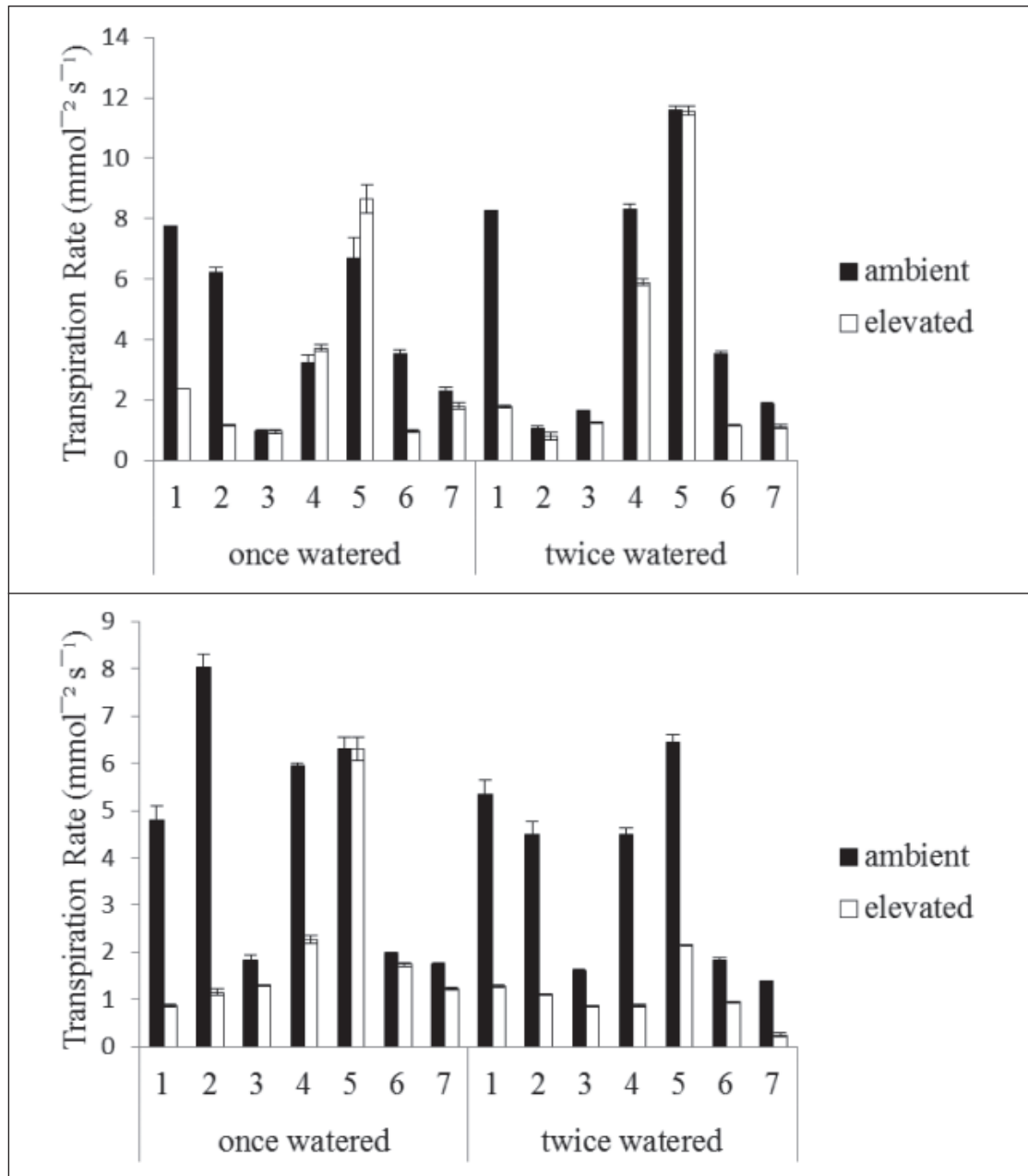


Fig. 7. Effect of elevated CO₂ concentration and water regime on the mean (\pm SE) transpiration rate of *M. malabathricum* (left) and *C. hirta* (right) ($p < 0.05$).

CO₂ ($p > 0.05$) (Figure 7). Our results differ than findings reported by Wang *et al* (2015), whom observed a higher rate of transpiration at elevated CO₂ on *Gynura bicolor*. The contrasting finding is possible as different types of plants react differently to elevated CO₂ and also water regime.

The highest value of **E** for *M. malabathricum* was observed in both elevated and ambient CO₂ and twice watered with the difference of **E** value of only 0.01 mmol⁻²s⁻¹ at the fifth month. The highest **E** for *M. malabathricum* plants in elevated CO₂ was on the fifth month and once watered with the value of 6.7 mmol⁻²s⁻¹. Meanwhile, *C. hirta* had the highest value

of **E** at ambient and once watered in the second month of treatment with the value of 8.0 mmol⁻²s⁻¹ followed by the one in ambient and twice watered with the value of 6.5 mmol⁻²s⁻¹ at the fifth month.

A lower rate of transpiration for the plants in elevated CO₂ compared to the ambient CO₂ may due to the diffusive resistance of H₂O is higher compared to the diffusive resistance of CO₂ and directly demonstrated that the plants in elevated CO₂ allowed the gas to diffuse into the leaves while they prevent H₂O from being released in order to protect themselves from dehydration (Drake *et al.*, 1997). Thongbai *et al* (2010) also pointed out that

transpiration will decrease as the CO₂ concentration increased due to an increased in air circulation. This means that transpiration rate and rate of stomatal conductance do affect each other at high CO₂ concentrations.

CONCLUSIONS

Positive effect of elevated CO₂ was more prominent for *M. malabathricum* compared to *C. hirta*. Morphologically, the positive effect of elevated CO₂ was exhibited by significant increase of plant height, below ground biomass and leaf area index of *M. malabathricum*. Whereas, for *C. hirta*, the positive effect of elevated CO₂ was seen in smaller increment of above ground biomass (instead of below ground biomass) and leaf area index. Both shrub species was morphologically not affected by water stress possibly due to their adaptation to extreme environment in urban areas. However, an interaction between CO₂ and water regime had the most significant effect on the leaf area index of both species.

The physiological responses of the two shrub species indicated that they are able to take advantage on elevated CO₂ concentration in urban areas, particularly for *M. malabathricum* and to a lesser extent for *C. hirta*. The different strategy used by the two species to cope with increased CO₂ may lies on the different natural microhabitat of these two species. *M. malabathricum* is a light-demanding shrub whereas *C. hirta* thrived better under shady habitat. It is also possible that confounding effects of other environmental factors such as light intensity may limit the two shrub plants response to elevated carbon dioxide. Therefore, we suggest other important growth limiting factor of different species should be considered in future elevated CO₂ experiment.

ACKNOWLEDGEMENTS

The authors would like to thank UKM Research Instrumentation Development Fund PIP-2013, Climate Change Institute (IKP) for providing the facilities of the experiment. The study was partly funded by FRGS/1/2014/STWN10/UKM/02/1. We would also like to thank kursi Perubahan Iklim UKM-YSD 27-2016-010 for funding the publication of this manuscript.

REFERENCES

- Ainsworth, E.A., Davey, P.A., Bernacchi, C.J. & Orla, C. 2002. A meta-analysis of elevated [CO₂] effects on soybean (*Glycine max*) physiology, growth and yield. *Global Change Biology*, **8**: 695-709.
- Ainsworth, E.A. & Long, S.P. 2005. What have we learned from 15 years of free-air CO₂ enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy properties and plant production to rising CO₂. *New Phytologist*, **165**(2): 351-372.
- Allen Jr, L.H., Bisbal, E.C., Campbell, W.J. & Boote, K.J. 1990. Carbon dioxide effects on soybean developmental stages and expansive growth. *In Proceedings-Soil and Crop Science Society of Florida*, **49**: 124-131.
- Anderson, L.J. & Cipollini, D. 2013. Gas exchange, growth, and defense responses of invasive *Alliaria petiolata* (Brassicaceae) and native *Geum vernum* (Rosaceae) to elevated atmospheric CO₂ and warm spring temperatures. *American Journal of Botany*, **100**(8): 1544-1554.
- Barnett, J. 2001. *Security and Climate Change*. Tyndall Centre for Climate Change Research. Working Paper 7. <http://tyndall.ac.uk/sites/default/files/wp7.pdf>. 20 pp.
- Bastin, J.F., Barbier, N., Réjou-Méchain, M., Fayolle, A., Gourlet-Fleury, S., Maniatis, D., de Haulleville, T., Baya, F., Beeckman, H., Beina, D., Couteron, P., Chuyong, G., Dauby, G., Doucet, J.L., Droissart, V., Dufrene, M., Ewango, C., Gillet, J.F., Gonmadje, C.H., Hart, T., Kacali, T., Kenfack, D., Libalah, M., Malhi, Y., Makana, J.R., Pelissier, R., Ploton, P., Serckx, A., Sonke, B., Stevart, T., Thomas, D.W., De Canniere, C. & Bogaert, J. 2015. Seeing Central African forests through their largest trees. *Scientific Reports*, **5**: 13156.
- Ceulemans, R. & Mousseau, M. 1994. Tansley Review No. 71. Effects of elevated atmospheric woody plants. *New Phytologist*, **127**(71): 425-446.
- Corlett, R.T. 2016. The Impacts of Droughts in Tropical Forests. *Trends in Plant Science*, **21**(7): 584-593.
- Drake, B.G., González-meler, M.A. & Long, S.P. 1997. More efficient plants: A Consequence of rising atmospheric CO₂?. *Annual Review of Plant Physiology*, **48**: 609-39.

- Farquhar, G.D. & Sharkey, T.D. 1982. Stomatal conductance and photosynthesis. *Annual Review of Plant Physiology*, **33(1)**: 317-345.
- Farrar, J.F. & Gunn, S. 1996. *Effects of temperature and atmospheric carbon dioxide on source-sink relations in the context of climate change*. Marcel Dekker, New York.
- Fauset, S., Johnson, M.O., Gloor, M., Baker, T.R., Monteagudo, M.A., Brienen, R.J.W., Feldpausch, T.R., Gonzalez, G.L., Malhi, Y., Steege, H., Pitman, N.C.A., Baraloto, C., Engel, J., Petronelli, P., Andrade, A., Camargo, J.L.C., Laurance, S.G.W., Laurance, W.F., Chave, J., Allie, E., Vargas, P.N., Terborgh, J.W. & Ruokolainen, K. 2015. Hyperdominance in Amazonian forest carbon cycling. *Nature Communications*, **6**: 6857.
- Franca, F., Lago, E.L. & Marsden, P.D. 1996. Plants used in the treatment of leishmanial ulcers due to *Leishmania (Viannia) braziliensis* in an endemic area of Bahia, Brazil. *Revista da Sociedade Brasileira de Medicina Tropical*, **29(3)**: 229-232.
- Geisinger, D.R., Johnson, D.W. & Ball, J.T. 1995. Enriched atmospheric CO₂, and soil P effects on growth and ectomycorrhizal colonization of juvenile ponderosa pine. *Forest Ecology and Management*, **78**: 207-215.
- Geisinger, D.R., Johnson, D.W. & Ball, J.T. 1998. Atmospheric CO₂, enrichment and soil N fertility effects on juvenile ponderosa pine: Growth, ectomycorrhizal development, and xylem water potential. *Forest Ecology and Management*, **102**: 33-44.
- International Sustainability Unit. 2015. *Tropical Forests – A Review*. Page Bros Limited, London. 145 pp.
- Joffry, S.M., Yob, N.J., Rofiee, M.S., Affandi, M.M.R.M.M., Suhaili, Z., Othman, F., Akim, A.M., Desa, M.N.M & Zakaria, Z.A. 2012. *Melastoma malabathricum* (L.) smith ethno-medicinal uses, chemical constituents, and pharmacological properties: A review. *Evidence-based Complementary and Alternative Medicine*, **2012**: 1-48.
- Johnson, S.N., Barton, A.T., Clark, K.E., Gregory, P.J., Mcmenemy, L.S. & Hancock, R.D. 2011. Elevated atmospheric carbon dioxide impairs the performance of root-feeding vine weevils by modifying root growth and secondary metabolites. *Global Change Biology*, **17(2)**: 688-695.
- Kirschbaum, M.U.F. 2011. Does enhanced photosynthesis enhance growth? Lessons learned from CO₂ enrichment studies. *Plant Physiology*, **155(1)**: 117-124.
- Maestre, F.T., Bowker, M.A., Puche, M.D., Belen Hinojosa, M., Martinez, I., Garcia-Palacios, P., Castillo, A.P., Soliveres, S., Luzuriaga, A.L., Sánchez, A.M., Carreira, J.A., Gallardo, A. & Escudero, A. 2009. Shrub encroachment can reverse desertification in semi-arid Mediterranean grasslands. *Ecology Letters*, **12(9)**: 930-941.
- Masle, J. 2000. The effects of elevated CO₂ concentrations on cell division rates, growth patterns, and blade anatomy in young wheat plants are modulated by factors related to leaf position, vernalization, and genotype. *Plant Physiology*, **122**: 1399-1415.
- Morison, J.I. 1985. Sensitivity of stomata and water use efficiency to high CO₂. *Plant, Cell & Environment*, **8(6)**: 467- 474.
- Nagy, M., Ogawa, K. & Hagihara, A. 2000. Interactive effect of CO₂ enrichment and temperature on the photosynthesis of field-grown hinoki cypress (*Chamaecyparis obtusa*) branches. *Trees*, **14(5)**: 0282-0288.
- Niell, F.X. 2003. Photon- and carbon-use efficiency in *Ulva rigida* at different CO₂ and N levels. *Planta*, **218**: 315-322.
- Omar, S.N.C. 2012. Potentials of *Melastoma malabathricum* Linn. flower and fruit extracts as antimicrobial infusions. *American Journal of Plant Sciences*, **3(8)**: 1127-1134.
- Roberntz, P. & Stockfors, J. 1998. Effects of elevated CO₂ concentration and nutrition on net photosynthesis, stomatal conductance and needle respiration of field-grown Norway spruce trees. *Tree Physiology*, **18(4)**: 233-241.
- Robredo, A. & Usue, P. 2007. Elevated CO₂ alleviates the impact of drought on barley improving water status by lowering stomatal conductance and delaying its effects on photosynthesis. *Environmental and Experimental Botany*, **59**: 252-263.
- Rogers, H.H., Prior, S.A., Runion, G.B. & Mitchell, R.J. 1996. Root to shoot ratio of crops as influenced by CO₂. *Plant and Soil*, **187**: 229-248.
- Sasek, T. & Strain, B. 1989. Effects of carbon dioxide enrichment on the expansion and size of Kudzu (*Pueraria lobata*) Leaves. *Weed Science*, **37(1)**: 23-28.
- Schaffer, B., Street, S.W., Whiley, A.W. & Searle, C. 1999. Atmospheric CO₂ enrichment, root restriction, photosynthesis and dry-matter partitioning in subtropical and tropical fruit crops. *Hortscience*, **34(6)**: 1033-1037.

- Schlesinger, W.H., Reynolds, J.F., Cunningham, G.L., Huenneke, L.F., Jarrell, W.M., Virginia, R.A. & Whitford, W.G. 1990. Biological feedbacks in global desertification. *Science*, **247(8)**: 1043-1048.
- Shimono, H., Nakamura, H., Hasegawa, T. & Okada, M. 2013. Lower responsiveness of canopy evapotranspiration rate than of leaf stomatal conductance to open-air CO₂ elevation in rice. *Global Change Biology*, **19(8)**: 2444-2453.
- Taylor, G., Ranasinghe, S., Bosac, C., Gardner, S.D.L. & Ferris, R. 1994. Elevated CO₂ and plant growth: cellular mechanisms and responses of whole plants. *Journal of Experimental Botany*, **45**: 1761-1774.
- Thomas, J.F. & Harvey, C.N. 1983. Leaf anatomy of four species grown under continuous CO₂ enrichment. *Botanical Gazette*, **144(3)**: 303-309.
- Thongbai, P., Kozai, T. & Ohyama, K. 2010. CO₂ and air circulation effects on photosynthesis and transpiration of tomato seedlings. *Scientia Horticulturae*, **126(3)**: 338-344.
- Wang, M., Dong, C., Fu, Y. & Liu, H. 2015. Growth, morphological and photosynthetic characteristics, antioxidant capacity, biomass yield and water use efficiency of *Gynura bicolor* DC exposed to super-elevated CO₂. *Acta Astronautica*, **114**: 138-146.
- Wester, L.L. & Wood, H.B. 1977. Koster's curse (*Clidemia hirta*), a weed pest in Hawaiian forests. *Environmental Conservation*, **4(1)**: 35-41.
- Wong, W. 2008. *Melastoma malabathricum*: Too beautiful to be called a weed. *Green Culture Singapore Feature Article*, August: 1-7.
- Woodrow, I.E. 1994. Optimal acclimation of the C3 photosynthetic system under enhanced CO₂. *Photosynthesis Research*, **39(3)**: 401-412.
- Xu, S., Zhu, X., Li, C. & Ye, Q. 2014. Effects of CO₂ enrichment on photosynthesis and growth in *Gerbera jamesonii*. *Scientia Horticulturae*, **177**: 77-84.