LEAF EPIDERMAL CHARACTERISTICS OF MEDICINAL Eclipta prostrata (L.) L., Vernonia amygdalina Delile (ASTERACEAE) AND Clitoria ternatea L. (FABACEAE)

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ABSTRACT

A study on leaf anatomical characteristics was undertaken on three medicinal plants species, *Eclipta prostrata* (L.) L., *Vernonia amygdalina* Delile and *Clitoria ternatea* L. It is found that there are variables in leaf epidermal features such as anticlinal wall patterns, stomata types, trichome types, and the occurrence of secretory structures. Results showed that there is slight variation in anticlinal wall patterns, which is, sinuous on abaxial surfaces and straight to wavy on adaxial surfaces of *Eclipta prostrata*, straight to wavy on both abaxial and adaxial surfaces of *Vernonia amygdalina*, and sinuous on both abaxial and adaxial surfaces of *Clitoria ternatea*, two types of stomata, which is anomocytic in *Eclipta prostrata* and *Vernonia amygdalina*, and paracytic in *Clitoria ternatea*, six types of trichomes recorded such as simple multicellular (short stalk – blunted tip), simple multicellular (echinate ornamentation, pointed tip), irregular T-shaped glandular (short to long stalk – elongated terminal), capitate glandular (short stalk – unicellular terminal) and peltate glandular. Secretory cells were observed in two species, *Eclipta prostrata* and *Vernonia amygdalina*. The leaf of these three medicinal species has been widely used in herbal preparation, however, there are not enough information nor a comprehensive study was done on the leaf anatomy of these species. Therefore, this study aimed to provide additional knowledge and more detailed information of the epidermal characteristics of *Eclipta*, *Vernonia* and *Clitoria* and contributes to systematic data available.

Key words: Anatomy, Asteraceae, *Clitoria ternatea*, *Eclipta prostrata*, Fabaceae, leaf epidermal, medicinal plants, *Vernonia amygdalina*

INTRODUCTION

Family Asteraceae, formerly known as Compositae, is the second largest family of flowering plants (Cronquist 2001), represented by about 950 genera

and 23000 species (Gills 1988), widely distributed throughout the world except for Antarctica (Funk & Susanna 2009). The members of the family are mostly herbs, sometimes shrubs and a few are trees (Bremer 1994). While family Fabaceae is known as one of the largest family amongst Angiosperms. This plant is

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native to many African countries (Mukherjee *et al.*, 2008) but has been extensively cultivated in the humid lowland tropics of Asia, Africa, Pacific Islands and Americas, and widespread throughout South-East Asia (Staples 1992).

Two of the species, Eclipta prostrata and Vernonia amygdalina belong to the family Asteraceae. Eclipta prostrata, locally known as Urang-aring, is an annual or short-lived perennial herb, found all around the tropics, extending towards the warm temperate, mostly found at the cultivated areas, along with watercourses and roadsides, about 10-80 cm tall, with prostrate erect branches, and numerous small, white heads (Burkill 1966; Jukema et al., 1991). On the other hand, Vernonia amygdalina, locally known as bismillah (Malay) or bitter leaf (English), due to its bitter taste, is a perennial shrub, grows wild in tropical Africa, small tree, about 1-6 m tall. The species is commonly grown and cultivated for its bitter leaves which are used for vegetables and medicinal purposes. Clitoria ternatea, locally known as kacang telang (Malay) or Butterfly Pea (English), is a member of the family Fabaceae. The plant is a perennial herbaceous climber and known for its deep-blue solitary flowers, used in food colouring and also medicinal purposes.

According to Rudall (1992), leaf epidermal surfaces characteristics are persistent characters in taxa, therefore, it has been used as the subject for study more than any other leaf parts. Leaf macromorphology characters such as shape, type, margin, apex and venation alone are not enough to give a precise decision in taxonomic conclusions and it is necessary to study the micro characters of the leaves to help in diagnosis (Taia 2004). Moreover, there are not enough information nor a comprehensive study was done on the leaf anatomy of these three medicinal species. Therefore, this study was designed to investigate and provide more detailed information and understanding of the epidermal characteristics and contributes to systematic data of genera Eclipta, Vernonia and Clitoria.

MATERIALS AND METHODS

Sources of plant specimens

Fresh specimens of *Eclipta prostrata, Vernonia amygdalina* and *Clitoria ternatea* with three replicates each, were collected from plots in the Malaysian Agricultural Research and Development Institute (MARDI) Serdang, Selangor. The specimens collected were authenticated by the curator/plant taxonomist at MARDI and the voucher specimens were deposited at the MDI Herbarium of the same institute.

Epidermal peels

Epidermal peels of both abaxial and adaxial surfaces of the three species were prepared by following a modified method of Johansen (1940) and Saas (1958). Pieces of 1 cm² of the leaves of each specimen were cut and soaked in Jeffery's fluid (10% nitric acid and 10% chromic acid, 1:1) in well-covered Petri dishes, for about four to six hours (depends on the sample) to macerate the mesophyll. The epidermis will be transferred into clean Petri dishes contain distilled water, then the abaxial and adaxial layers will be separated using a fine brush and forceps. The isolated epidermal layers then will be rinsed and cleansed with distilled water. Afterwards, the tissues were stained with Safranin and Alcian Blue solution for about five minutes each and then rinsed with distilled water to remove excess staining and transferred to a clear glass microscopic slide. The slides were then observed under a Fisher light microscope with ×20 and ×40 magnifications. Photographs of the epidermal peels were taken using Olympus SZH40 microscope (Tokyo, Japan) and images were processed using two software, Image Analysis and Adobe Photoshop. After observations, permanent slides were prepared. The tissues were then undergone dehydration process, using 50%, 70%, 95% and 100% alcohol, and mounted on the slides using Euparal, covered with coverslips and placed in the oven at 60°C for nearly two weeks, before being kept in box slides for future references. Three slides were prepared for each abaxial and adaxial epidermal peels of the three species. Table 1 showed steps for staining, dehydration, mounting and drying of epidermal peels.

Analysis and descriptions

Details of the analysis and descriptions on epidermal anatomy characteristics followed those of Metcalfe and Chalk (1965).

RESULTS AND DISCUSSION

The epidermal cells and stomatal characteristics of the three species studied are summarized in Table 2 and illustrated in Figure 1, 2 and 3.

Anticlinal walls

The anticlinal wall in *Eclipta prostrata* is sinuous on abaxial surface, and straight to wavy on the adaxial surface (Figure 1), straight to wavy on both abaxial and adaxial surfaces of *Vernonia amygdalina* (Figure 2), and sinuous on both abaxial and adaxial surfaces of *Clitoria ternatea* (Figure 3). These results supported previous study on these species, that undulate-cuneate or sinuous anticlinal

Table 1. Steps for staining, dehydration, mounting and drying process

Step	Solution	Time	Notes
Staining			
1	2–3 drops of Safranin into petri dish contains the tissue and distilled water	5 mins.	-
2	Rinsed with distilled water. Repeat three times	_	_
3	2-3 drops of Alcian Blue into the Petri dish contains the tissue and distilled water	5 mins.	-
4	Rinsed with distilled water. Repeat three times	-	_
Dehydration			
1	Alcohol 50%	5 mins.	Close petri dish
2	Alcohol 70% (+ a drop of hydrochloric acid- HCl)	2 mins.	Close and stir petri
3	Alcohol 95%	5 mins.	Close petri dish
4	Alcohol 100%	5 mins.	Close petri dish
Mounting			
3	Euparal	-	
Drying			
	-	2 weeks	Placed in the oven at 60°C

Table 2. Epidermal cells and stomatal characteristics of species studied

Species	Epidermal surface	Anticlinal walls	Stomatal type	Trichome type	Secretory cells occurrence
Eclipta prostrata	Abaxial	Sinuous	Anomocytic	Capitate glandular (short stalk – unicellular terminal), Simple multicellular (echinate ornamentation, pointed tip)	Present
	Adaxial	Straight to wavy	Anomocytic	Simple multicellular (echinate ornamentation, pointed tip), Simple multicellular (short stalk – blunted tip)	Absent
Vernonia amygdalina	Abaxial	Straight to wavy	Anomocytic	Irregular T-shaped glandular (short to long stalk – elongated terminal), Peltate glandular	Absent
	Adaxial	Straight to wavy	Anomocytic	Simple multicellular (short to long stalk – blunted tip), Peltate glandular	Present
Clitoria ternatea	Abaxial	Sinuous	Paracytic	Simple multicellular (echinate ornamentation, pointed tip),	Absent
	Adaxial	Sinuous	Paracytic	Simple multicellular (echinate ornamentation, pointed tip)	Absent

walls occurred in *Clitoria ternatea* (Khatijah *et al.*, 2006; Chukwuma *et al.*, 2014) and straight-arcuate or undulating anticlinal walls occurred in *Vernonia amygdalina* (Adedeji & Jewoola, 2008; Kemka *et al.*, 2014). The anticlinal wall of *Eclipta prostrata* is reported for the first time.

Stomata types

The distribution of the stomata in all three species is amphistomatic, which is stomata occurring on both abaxial and adaxial surface (Table 2).

Anomocytic stomata were found to occur in *Eclipta* prostrata and *Vernonia amygdalina* (Figure 1B, 1F, 2B and 2F), while paracytic stomata occurred in *Clitoria ternatea* (Figure 3A and 3D). This result supports Metcalfe and Chalk (1965), which reported that stomata in Compositae (Asteraceae) is generally 'ranunculaceous or anomocytic, and 'rubiaceous' or paracytic in Leguminosae (Fabaceae). Perveen *et al.* (2007) in a study on stomatal types of dicots, also reported that anomocytic stomata occurred in *Eclipta prostrata*. Chukwuma *et al.* (2014) reported

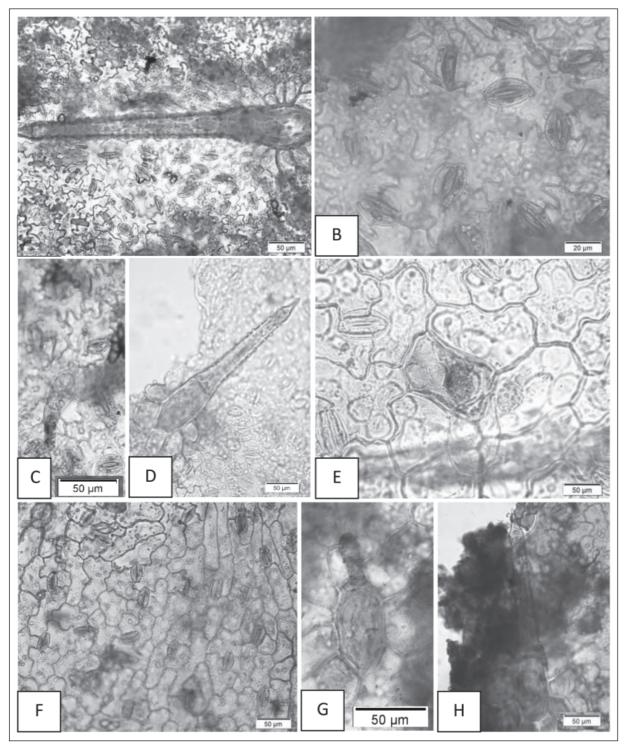


Fig. 1. Leaf epidermal of *Eclipta prostrata*. (A-E) Abaxial leaf surface. A) Sinuous anticlinal walls, B) Anomocytic stomata, C) Capitate glandular (short stalk – unicelular terminal), D) Simple multicellular (echinate ornamentation, pointed tip), E) Secretory cells. (F-H) Adaxial leaf surface. F) Straight to wavy anticlinal walls with anomocytic stomata, G) Simple multicellular (short stalk – blunted tip), H) Simple multicellular (echinate ornamentation, pointed tip). Scale: A, C-H) 50 μm, B) 20 μm.

that there are three types of trichome present in *Clitoria ternatea*, which is paracytic, anisocytic and anomocytic, however, this study only found paracytic stomata.

Trichome types

There are five trichome types observed in the study, including both non-glandular and glandular types (Table 2). Non-glandular trichomes are

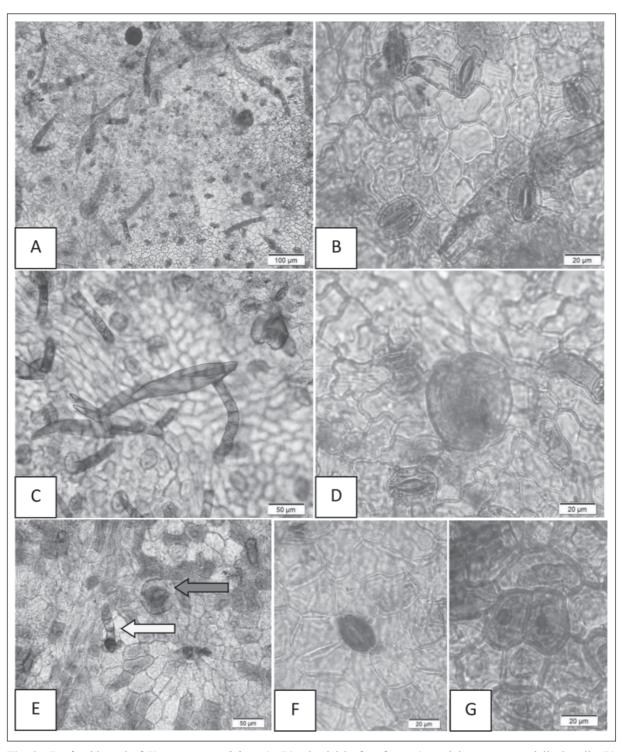


Fig. 2. Leaf epidermal of *Vernonia amygdalina*. (A-D) Abaxial leaf surface. A) Straight to wavy anticlinal walls, B) Anomocytic stomata, C) Irregular T-shaped glandular (short to long stalk – elongated terminal), D) Peltate glandular. (E-G) Adaxial leaf surface. E) Straight to wavy anticlinal walls with simple multicellular (short stalk – blunted tip), (yellow arrow) and peltate glandular (red arrow) trichomes, F) Anomocytic stomata, G) Secretory cells. Scale: A) 100 μ m, C & E) 50 μ m, B, D, F & G) 20 μ m.

simple multicellular (short stalk – blunted tip), simple multicellular (echinate ornamentation, pointed tip), and simple unicellular (echinate ornamentation, pointed tip), while glandular trichomes found are irregular T-shaped glandular (short to long stalk –

elongated terminal), capitate glandular (short stalk – unicellular terminal) and peltate glandular.

Three types of trichomes were found in *Eclipta prostrata*, capitate glandular (short stalk – unicellular terminal) (Figure 1C), simple multicellular (echinate

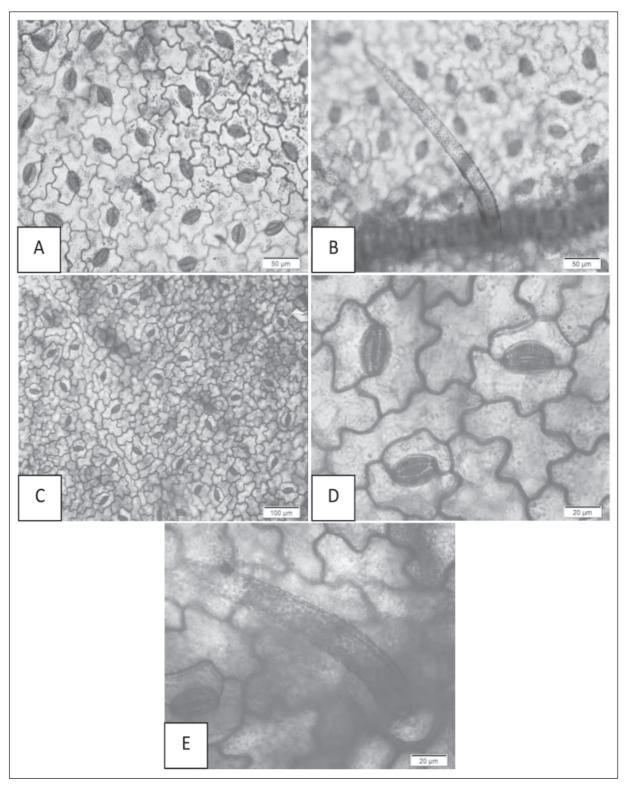


Fig. 3. Leaf epidermal of *Clitoria ternatea*. (A-B) Abaxial leaf surface. A) Sinuous anticlinal walls with paracytic stomata, B) Simple unicellular (echinate ornamentation, pointed tip), (C-E) Adaxial leaf surface. C) Sinuous anticlinal walls, D) Paracytic stomata, E) Simple unicellular (echinate ornamentation, pointed tip). Scale: A & B) 50 μm, C) 100 μm, D & E) 20 μm.

ornamentation, pointed tip) (Figure 1D), and simple multicellular (short stalk – blunted tip) (Figure 1G). Shafira and Salamah (2020) also reported the occurrence of simple multicellular (echinate

ornamentation, pointed tip) and simple multicellular (short stalk – blunted tip) on leaves of *Eclipta prostrata* in their study on trichomes of four species in the family Asteraceae. This is the first time capitate

glandular (short stalk – unicellular terminal) trichome reported for the species.

Three types of trichomes were also recorded in *Vernonia amygdalina*, which are, irregular T-shaped glandular (short to long stalk – elongated terminal) (Figure 2C), peltate glandular (Figure 2D), and simple multicellular (short stalk – blunted tip) (Figure 2E). This information supports the previous study done by Oladele (1990) and Kemka *et al.* (2014). They stated that the occurrence and morphology of the irregular T-shaped glandular and bilobed glandular trichome (peltate glandular) is a diagnostic character for *Vernonia amygdalina*. A simple multicellular (short stalk – blunted tip) trichome was also reported for the first time for *Vernonia amygdalina*.

Only one type of trichome was found *Clitoria ternatea*, which is simple unicellular (echinate ornamentation, pointed tip) (Figure 3B and 3E). Khatoon *et al.* (2015) also found the same trichomes in their pharmacognostical studies on blue and white flower varieties of *Clitoria ternatea*.

Occurrence of secretory structures

Castro et al. (1997) have mentioned that there are eight types of leaf secretory structures found in Asteraceae; ducts, cavities, idioblasts, laticifers, hydathodes, extrafloral nectaries, trichomes and glandular appendixes. According to Metcalfe and Chalk (1965), latex sometimes presents in elongated secretory cells in *Vernonia*, while the occurrence of secretory elements in Fabaceae can be in two ways, in cells or sacs.

Secretory structures were found in two species are, *Eclipta prostrata* and *Vernonia amygdalina*, both in ducts form and glandular trichomes. *Clitoria ternatea*, however, showed no occurrence of secretory structures. In *Eclipta prostrata*, the secretory cells are elongated, almost rectangular shape, while in *Vernonia amygdalina*, the cells are shorter and rounded. Lersten and Curtis (1987) also have discussed the wide variability in internal secretory structures of the Asteraceae. Despite their differences in positions and size, these secretory structures are usually filled with thick yellowish to a dark brown oil, in a form of spherical droplets.

CONCLUSIONS

This present finding has proved that leaf epidermal characteristics of *Eclipta prostrata*, *Vernonia amygdalina* and *Clitoria ternatea*, such as anticlinal wall patterns, stomata types, trichome types, and occurrence of secretory structures could serve useful in the identifications of these three medicinal plant species. While this work supports the previous study, there are also new findings reported,

contributed to the existing taxonomic and systematic information on the species studied and may be used in the identification of the plant species in the absence of its inflorescences. The anticlinal wall of *Eclipta prostrata* is reported for the first time in this study. Capitate glandular (short stalk – unicellular terminal) trichome and simple multicellular (short stalk – blunted tip) trichome were also reported for the first time for *Eclipta prostrata* and *Vernonia amygdalina* respectively. Although leaf epidermal characteristics have contributed greatly to the taxonomy of angiosperms, more studies need to be done to provide detailed information and expand the knowledge on the species studied.

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