ECOLOGY AND DISTRIBUTION OF MOSQUITO LARVAE IN THE INLAND HABITAT OF SOUTH SULAWESI, INDONESIA

Arini Ratnasari^{1,4*}, Isra Wahid², Hairil Akbar³, Agnes Immanuela Toemon⁴, Arif Rahman Jabal⁴, Dian Mutiasari⁵, Matius Paundanan⁶, Hanasia⁷, Awaluddin⁸, Sugiarto⁹ & M Ikhsan¹⁰ ¹Graduate School, Faculty of Medicine. Hasanuddin University, Makassar, South Sulawesi, Indonesia ²Faculty of Medicine, Universitas Hasanuddin. Jl. Perintis Kemerdekaan Km. 10. Makassar 902425, South Sulawesi, Indonesia ³Public Health Program, Graha Medika Institute of Health and Technology, Kotamobago, North Sulawesi, Indonesia ⁴Departement of Parasitology, Faculty of Medicine, Palangka Raya University, Palangka Raya, Central Kalimantan, Indonesia ⁵ Departement of Public Health, Faculty of Medicine, Palangka Raya University, Palangka Raya, Indonesia ⁶ Public Health Program, Indonesia Jaya Institute of Health Science, Palu, Central Sulawesi, Indonesia ⁷ Medical Program, Faculty of Medicine, Palangka Raya University, Palangka Raya, Central Kalimantan, Indonesia ⁸ Medical Laboratory Technology Program, Megarezky University, Makassar, South Sulawesi, Indonesia ⁹Ministry of Health Indonesia, South Jakarta, Jakarta, Indonesia ¹⁰Study Program of Parasitology & Medical Entomology, Bogor Agricultural University, Bogor, West Java, Indonesia *Corresponding author: ariniratnasari.ento@gmail.com

Submission: 18 April 2022; Acceptance: 19 February 2023

ABSTRACT

The ecology and distribution of mosquito larvae have a significant effect on mosquito populations and in arbovirus transmission. There is no information on the ecology and distribution of mosquito larvae in the inland area of South Sulawesi, Indonesia. This study aimed to examine distribution and ecological of mosquito larvae that affect the survival of arbovirus vectors in the inland area of South Sulawesi. Larvae were obtained in eight locations of the South Sulawesi region. The collected samples were examined in the Entomology Laboratory, Faculty of Medicine, Hasanuddin University, South Sulawesi, Indonesia. Larvae were counted, separated by genus and identified to the species level. The physical parameters of the water in the breeding sites (pH, temperature and salinity) and the ambient relative humidity were recorded. Data were analyzed using IBM SPSS version 24 and the spatial distribution of larvae was mapped using ArcGIS version 10.5. Results showed that there were 3,803 larvae consisting of Ae. aegypti, Ae. albopictus, Cx. quinquefasciatus, Cx. vishnui, Cx. tritaenorynchus, Cx. gelidus, and An. vagus. The physical parameters of the water habitat for mosquito larvae range from a temperature of 24.5°C-34.2°C, water pH 6.3-7.5, humidity 67.8-89.0, and salinity 0 ppt. We found that mosquito larvae abundant had a significant relationship (P=0.00) with pH, salinity, temperature, and humidity. Results suggest that the abundance of arbovirus vectors in the inland of South Sulawesi can be controlled by vector control programs.

Keywords: Arbovirus, mosquito, inland area, South Sulawesi

ABSTRAK

Ekologi dan taburan larva nyamuk mempunyai kesan signifikan ke atas populasi nyamuk dalam transmisi arbovirus. Masih tiada maklumat ke atas ekologi dan taburan larva nyamuk ini di pedalaman Sulawesi Selatan, Indonesia. Kajian ini bertujuan untuk menilai taburan dan ekologi larva nyamuk bagi menilai jangka hayat vektor arbovirus di pedalaman Sulawei Selatan, Indonesia. Sampel yang dikumpul telah diperiksa di Makmal Entomologi, Fakulti Perubatan, Hasanuddin University, Selatan Sulawesi, Indonesia. Larva dikira, diasingkan ke peringkat genus dan dicamkan ke peringkat spesies. Parameter fizikal untuk air yang digunakan untuk tempat pembiakan (pH, suhu dan kemasinan) dan kelembapan relatif ambien telah direkodkan. Data telah dianalisis menggunakan IBM SPSS version 24 dan taburan ruang larva diplotkan menggunakan ArcGIS versi 10.5. Hasil menunjukkan sejumlah 3,803 larva merangkumi Ae. aegypti, Ae. albopictus, Cx. quinquefasciatus, Cx. vishnui, Cx. tritaenorynchus, Cx. gelidus, dan An. vagus. Parameter fizikal pada habitat air untuk larva nyamuk suhunya di 24.5°C-34.2°C, pH air 6.3-7.5, kelembapan 67.8-89.0 dan kemasinan pada 0 ppt. Kelimpahan larva nyamuk menunjukkan hubungan yang signifikan (P=0.00) ke atas faktor pH, kemasinan, suhu, dan kelembapan. Kelembapan vektor arbovirus di pedalaman Selatan Sulawesi dapat dikawal melalui program kawalan vektor.

Kata kunci: Arbovirus, nyamuk, kawasan pedalaman, Selatan Sulawesi

INTRODUCTION

Mosquitoes are vectors of arboviruses and parasites such as malaria, chikungunya, dengue fever, and Japanese encephalitis (Ratnasari et al. 2020). In South Sulawesi, the acute infection of malaria, chikungunya, dengue hemorrhagic fever, and Japanese encephalitis were 0.13, 7, 19.96, and 52.3%, respectively (Arif et al. 2020; Ministry of Health Indonesia 2018; Widarso

et al. 2002). Arboviruses and parasites transmitted by mosquitoes are associated with mosquito life-supporting habitats, one of which is inland areas far from the coast.

The inland portion of South Sulawesi is primarily used as agricultural land and settlements by the locals, and both places support mosquito habitats. According to Hamidun et al. (2021) ecological habitat of mosquito larvae in residential area and landscaping nursery area. Mosquito larvae can be found in forests and non-forest (Khariri, 2018). Mosquito larvae can be found on the banks of rivers (Chaiphongpachara et al. 2018). Mosquito larvae were related humadity, rainfall, dan temperature, precipitation (Drakou et al. 2020; Kesetyaningsih et al. 2018).

Larvae of *Anopheles* sp. can be found in rice fields, puddles, and irrigation canals (Inunggita et al. 2019; Robert et al. 2002). According to Ratnasari et al. (2021), the breeding places of *Ae. aegypti* can be found in bathrooms, used buckets, old tires, flowerpots, trash cans, paint cans, and water tanks. Meanwhile, Augustina et al. (2021) found that the habitat of *Culex* sp. was in used pots, swamps, sewers, and canals.

Ecological surveys of arbovirus and parasitic vector habitats are ineffective in inland areas due to various natural and artificial mosquito breeding sites. Therefore, many different mosquito habitats pose challenges to controlling and eradicating arboviruses and parasites. In addition, there is no information on the ecology and distribution of mosquito larvae in the inland area of South Sulawesi. The aim of the study analyzed mosquito larvae in inland areas, which have the potential as arbovirus and parasite vectors.

MATERIALS AND METHODS

This observational study used simple random sampling to collect the distribution of mosquito larvae and habitat ecology in the inland region of South Sulawesi. Simple random sampling is a random sampling technique without distinguishing those in the population.

Study Areas

Mosquito larvae were collected in inland South Sulawesi from January 2018 to December 2019. Should be eight districts include the district of this city were chosen for sampling (Table 1). The location is chosen based on the ease of access and the number of dengue cases (Figure 1).

Indonesia			
District/City	Sub-District	Lattitude	Longitude
Makassar	Biringkanaya	5°03'35.6" S	119°28'8.37" E
Maros	Marusu	5°03'71.5" S	119°49'91.6" E
Gowa	Pattalassang	5°12'32.3" S	119°33'26.5" E
Takalar	Palombangkeng	5°24'14.0" S	119°30'02.9" E
Jeneponto	Bontoramba	5°35'28.0" S	119°40'06.2" E
Bantaeng	Sinoa	5°29'54.8" S	119°55'27.3" E
Bulukumba	Rilau Ale	5°26'34.9" S	120°12'24.0" E
Pangkep	Balocci	4°53'49.4" S	119°38'13.2" E

 Table 1.
 Latitudes and longitudes of the study locations in the inland area of South Sulawesi, Indonesia



Figure 1. Map showing the eight districts of the inland area of South Sulawesi with the selected sampling location shown with pin marker

Insect Sampling

Sampling was carried out from morning to noon (8:00 a.m. to 12:00 p.m. local time) by searching for breeding habitats for mosquito larvae. A Pasteur pipette with 10 cm diameter filter was used for small samples, whereas a dipper was used for large numbers of larvae. In addition, all mosquito larvae in containers with water were transferred to sample bottles, labelled, and the water's physical parameters were measured. The coordinates for each location were recorded using a Garmin GPS eTrex-10 series. Collected samples were brought to the Entomology Laboratory of Hasanuddin University for identification and analysis. A sampling at eight locations based on variations in densely populated inland areas rarely touched by mosquito eradication programs. Consideration of location selection based on easy access to span that has high data on cases of dengue hemorrhagic fever.

Physical Parameters Data Collection

Measurement of physical parameters in larval samples included pH, temperature, humidity, and salinity using digital tools. The parameters of pH and temperature were measured using an instrument (Yieryi-3508), a salinometer (Lutron YK-31SA), and a hygrometer (AMF051).

Species Identification

Mosquito larvae were counted and grouped based on the sampling location. The mosquito larvae were identified using a stereomicroscope and taxonomic identification key by Rueda (2004), O'Connor and Soepanto (1999) and Ministry of Health Indonesia (2017).

Data Analysis

The chi-square analysis assessed the number and percentage of larvae in each habitat. The Pearson correlation test examined the relationship between the number of larvae in each location and physical parameters. Finally, the spatial distribution of larvae was mapped using the ArcGIS 10.5 application.

RESULTS AND DISCUSSION

A total of 3,803 larvae were collected during field observations consisting of three genera and seven species with include *Ae. aegypti* (n=2,059), *Ae. albopictus* (n=540), *Cx. quinquefasciatus* (n=768), *Cx. vishnui* (n=118), *Cx. tritaeniorhynchus* (n=233), *Cx. gelidus* (n=6), and *An. vagus* (n=79) show in Table 2 and Figure 2.

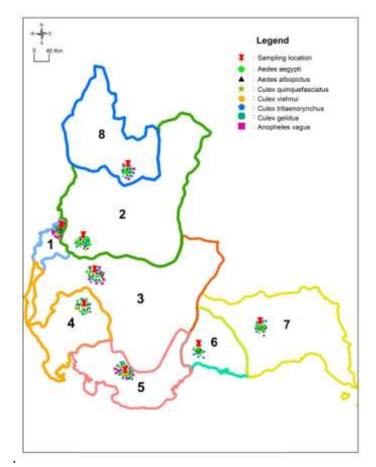


Figure 2. Map of distribution larvae in the inland area of South Sulawesi

The species distribution and numbers of mosquito larvae collected from breeding sites in inland areas of South Sulawesi of mosquito are shown in Table 2 and Figure 2. The larvae breeding habitats preference shows variation between genus and species. Table 3 and Figure 3 show four types of containers with high larval percentages, i.e., used buckets (33.3%), used tires (26.4%), water drums (17.2%), and bathtubs (12.2%).

Ratnasari et al.

Location	Ae. aegypti	Ae. albopictus	Cx. quinquefasciatus	Cx. vishnui	Cx. tritaenorynchus	Cx. gelidus	An. vagus	Total
Biringkanaya	345	189	176	23	76	0	16	825
Marusu	276	102	122	13	45	0	0	558
Pattalassang	177	48	98	0	33	0	39	395
Palombangkeng	197	72	73	3	28	1	0	374
Bontoramba	110	29	102	78	18	5	21	363
Sinoa	293	52	16	0	8	0	0	369
Rilau Ale	210	11	54	1	21	0	0	297
Balocci	451	37	127	0	4	0	3	622
Total	2059	540	768	118	233	6	79	3803

Table 2.Species and numbers of mosquito larvae collected from breeding sites ininland areas of South Sulawesi

 Table 3.
 Types of containers and percentage of mosquito larvae collected in the inland area of South Sulawesi

Tupo of	Total	Number of	Larvae Percentage (%)						
Type of containers	number of containers	Positive Container	Ae. aegypti	Ae. albopictus	Cx. quinquefasciatus	Cx. vishnui	Cx. tritaenorynchus	Cx. gelidus	An. vagus
Used tires	543	311	26.4	17.2	8.7	0	0	0	0
Flowerpot	77	36	6.4	2.0	2.5	0	0	0	0
Plastic cups	160	26	2.2	0	0.1	0	0	0	0
Used Bucket	381	128	10.7	5.9	11.6	0	0	0	0
Used Pans	48	16	5.0	10.7	2.0	0	0	0	0
Used Gallons	33	12	4.7	0	0	0	0	0	0
Trash can	20	9	3.7	4.8	7.3	0	0	0	0
Petals	10	6	0.5	3.3	0	0	0	0	0
Water hole	8	3	0	0	0.4	0	16.0	0	7.3
Irrigation Channel	30	9	0	0	22.1	8.6	12.0	0	13.0
Rice field	10	5	0	0	11.2	22.0	8.0	32.8	79.7
Swamp	8	6	0	0	18.9	54.0	64.0	67.2	0

Serangga 2023, 28(1): 128-145.

Ratnasari et al.

Total	2320	1095	100	100	100	100	100	100	100
Crock	12	3	3.0	4.3	10.2	0	0	0	0
Tree hole	28	7	0	16.4	0	0	0	0	0
Dispenser	47	20	2.6	0	0	0	0	0	0
Soil pitcher	56	15	1.4	0	0	0	0	0	0
Bath	293	137	7.4	12.2	0	0	0	0	0
Used dipper	23	3	0.4	0.4	0	0	0	0	0
Pool	7	3	0.6	0.9	0	15.4	0	0	0
Styrofoam	87	17	2.3	1.7	0	0	0	0	0
Used cans	63	34	8.6	3.9	1.0	0	0	0	0
Water drums	376	289	14.0	16.3	4.0	0	0	0	0



Figure 4. Mosquito breeding places: a. used tires, b. water drum, c. used bucket, d. bath, e. irrigation channel, f. water hole, g. swamp, h. ricefield

ISSN 1394-5130

The physical properties of water influence the breeding, survival, and adaptation of various organisms, including mosquito larvae. Mosquitoes can breed in various aquatic environments such as in abandoned containers (used tires, water drums and buckets), natural containers (water holes and rock pools) and water bodies with zero salinity (irrigation channels and rice fields). Mosquito larvae require water temperatures ranging from 24.5°C to 34.2°C, water pH ranging from 6.3 to 7.5, and humidity ranging from 67.8 to 89.0 (Table 4).

Table 4.Physical properties of mosquito larvae habitat in the inland area of South Sulawesi							
Location	pН	Salinity (ppt)	Temperature (°C)	Humidity (%)			
Biringkanaya	±6.7-7.2	0	$\pm 27.3 - 32.4$	$\pm 70.3 - 85.0$			
Marusu	$\pm 6.3-7.0$	0	$\pm 24.5 - 33.2$	$\pm 69.8 - 82.8$			
Pattalassang	$\pm 6.9-7.5$	0	$\pm 26.9 - 32.0$	$\pm 73.2 - 84.2$			
Palombangkeng	$\pm 7.0-7.3$	0	$\pm 25.3 - 31.4$	$\pm 70.5 - 86.4$			
Bontoramba	$\pm 6.8-7.4$	0	$\pm 28.0-34.2$	$\pm 67.8 - 89.0$			
Sinoa	±6.6-7.1	0	$\pm 28.5 - 32.5$	$\pm 70.8 - 82.4$			
Rilau Ale	$\pm 7.0-7.4$	0	$\pm 26.2 - 31.0$	$\pm 71.0-88.2$			
Balocci	$\pm 7.0-7.2$	0	$\pm 28.2 - 34.2$	$\pm 69.7 - 86.4$			

Chi-Square analysis showed significant habitat characteristics with an abundance of *Ae. aegypti, Ae. albopictus, Cx. quinquefasciatus, Cx. vishnui, Cx. tritaeniorhynchus*, while in *Cx. gelidus* and *An. vagus* was not significantly associated ($P \ge 0.05$). The number of mosquito larvae found was related to pH, salinity, temperature, and humidity (Table 5).

0	Chi-Square Tests					
Spesies		Value	Df	Sig.		
Ae. aegypti	Pearson Chi-Square	16.336	20	0.023*		
	Likelihood Ratio	17.602	20	0.044		
	Linear-by-Linear Association	0.489	1	0.184		
Ae. albopictus	Pearson Chi-Square	13.080	20	0.034*		
	Likelihood Ratio	15.134	20	0.514		
	Linear-by-Linear Association	0.889	1	0.346		
Cx. quinquefasciatus	Pearson Chi-Square	13.753	20	0.019*		
	Likelihood Ratio	15.079	20	0.418		
	Linear-by-Linear Association	1.631	1	0.202		
Cx. vishnui	Pearson Chi-Square	21.342	20	0.031*		
	Likelihood Ratio	26.326	20	0.121		
	Linear-by-Linear Association	0.845	1	0.158		
Cx. tritaenorynchus	Pearson Chi-Square	14.425	20	0.025*		

Table 5.Habitat characteristics correlated with the abundance of the mosquito larvae in
inland area

	Likelihood Ratio	15.259	20	0.006
	Linear-by-Linear Association	2.221	1	0.136
Cx. gelidus	Pearson Chi-Square	10.753	20	0.932
	Likelihood Ratio	12.014	20	0.885
	Linear-by-Linear Association	1.055	1	0.304
An. vagus	Pearson Chi-Square	18.525	20	0.488
	Likelihood Ratio	19.618	20	0.418
	Linear-by-Linear Association	0.093	1	0.760

Note: *Correlation is significant at the 0.05 level (2-tailed)

Most of the vectors of arboviruses in humans from the Culicidae family belong to the phylum Arthropoda. Arboviruses are found in tropical and sub-tropical countries, including Indonesia (Myint et al. 2014). Arbovirus infections have made up 30% of all infectious diseases (Jones et al. 2008). *Aedes* sp. lives in tropical and subtropical areas, breeds in plastic containers, tires, pots, leaf petals and can adapt to various environments (Ferede et al. 2018). *Ae. aegypti* and *Ae. albopictus* are vectors of dengue virus (Guo et al. 2016; Ibáñez-Justicia et al. 2020; Satoto et al. 2014).

Breeding places for *Culex* sp. were found in puddles of water, rice fields, irrigation canals, used tires, ditches, soil holes, and unused ponds (Nchoutpouen et al. 2019; Shaman 2010). The abundance of *Culex* sp. mosquitoes has the potential to pose a risk of spreading Japanese encephalitis (Lindahl et al. 2012), Zika (Huang et al. 2016), Usutu, and West Nile viruses (Fros et al. 2015). In addition, *Culex* sp. that is a carrier of the West Nile virus include *Cx. tritaeniorhynchus*, *Cx. pipiens*, *Cx. nigrapalpus*, *Cx. quinquefasciatus*, *Cx. restuans*, and *Cx. modestus* (Golding et al. 2012).

Anopheles sp. is a carrier of malaria that causes the O'nyong-nyong virus in humans (Rezza et al. 2017). Currently, malaria is still a serious health problem in Indonesia. The high risk of death occurs in infants, children under five years, and pregnant women (Jiero & Pasaribu 2021; Patriani et al. 2019). Anopheles habitats were found in larger water bodies such as lakes, river banks, swamps, ponds, rice fields, and rivers (Hamidian 2011). The early discovery of breeding sites for arbovirus-carrying mosquito larvae is one of the strategies to prevent the spread of arboviruses.

Distribution of larvae of *Ae. aegypti* in this study was found in Balocci, *Ae. albopictus*, *Cx. tritaenorynchus* and *Cx. quinquefasciatus* in Biringkanaya, *Cx. vishnui* and *Cx. gelidus* in Bontoramba, and *An. vagus* in Pattalassang. The abundance of mosquito larvae in the inland area of each site is related to the proximity of larval habitat containers from residential areas. Based on Augustina et al. (2021), mosquito larvae are found to breed mostly in old tires, puddles, plastic containers, pots, and drains in inland areas. Vector abundance is related to residential areas (Rahma et al. 2020). Characteristics of mosquito larvae breeding sites in the inland areas were found in various containers. *Ae. aegypti* and *Ae. albopictus* was found highest in used tires, *Cx. quinquefasciatus* in irrigation canals, *Cx. vishnui*, *Cx. tritaenorynchus*, and *Cx. gelidus* in swamps, as well as *An. vagus* in the rice fields.

Larvae of *Ae. aegypti* and *Ae. albopictus* is often found in used tires because people store used tires in their yards. People use old tires as artificial pots and waste storage. The

disadvantages of using used tires is that they collect rain water, and when neglected, *Ae. aegypti* and *Ae. albopictus* tend to lay eggs in these tires. A study by Ferede et al. (2018) have shown a significant percentage (57.5%) of *Ae. aegypti* breeding in used tires. Other studies have shown a substantial percentage of mix breeding of *Ae. albopictus* and *Ae. aegypti* in used tires ie; 45% (Futami et al. 2020) 26.5% (Higa et al. 2010) and 9.1% (Ratnasari et al. 2021).

Larvae of *Cx. quinquefasciatus* is commonly found in irrigation canals filled with garbage. They Mosquitoes will lay their eggs in slow-flowing, calm, and litter-filled water. *Cx. quinquefasciatus* is widespread in urban, suburban, rural, and remote areas. These mosquitoes are often found in various habitats and are anthropophilic (Muturi et al. 2007; Reuben 1992). The distribution of *Cx. quinquefasciatus* is found in various countries such as Thailand (Kitvatanachai et al. 2005), Singapore (Lam-Phua et al. 2019), Philippines (Carvajal et al. 2018), Vietnam (Ha et al. 2021) and Malaysia (Low et al. 2012; Ng et al. 2016).

Larvae of *Cx. vishnui*, *Cx. tritaenorynchus*, and *Cx. gelidus* were found in irrigation canals, rice fields, and most larvae were found in swamps. Larvae of *Cx. vishnui* and *Cx. gelidus* can be found in lowland and mountainous areas. Habitat characteristics of these two species are found in rice fields, cultivated areas, and livestock. The lowest percentage of *Cx. gelidus* larvae in this study were influenced by residents who did not own livestock such as pigs. *Cx. gelidus* prefer to suck the blood of animals such as pigs and human blood (Lindahl et al. 2012).

The percentage of larvae of *An. vagus* was found in water holes and irrigation channels. The highest number larvae of *An. vagus* was found in rice fields. Some of the villagers are exposed to *An. vagus* because they have rice fields near their houses which support this mosquito to breed. A study by Wharton et al. (1963) found that breeding places for Anopheles were located in swamps. In addition, several potential places include rice fields, waterways, buffalo puddles, and rivers (Maretasari et al. 2019).

The physical factor of water is very influential in the success of breeding and survival of mosquito larvae. These factors can affect the density and diversity of mosquitoes. Water quality and characteristics are determinants of species diversity and mosquito composition (Bashar et al. 2016). This study showed that the water in the container had varying pH, temperature, humidity, and salinity, which could affect the survival of the larvae. A total of eight locations showed that pH, temperature, and humidity supported the breeding of *Aedes*, *Culex*, and *Anopheles*. According to Lubinda et al. (2019) and Chandrasegaran et al. (2020), the larvae of *Ae. aegypti* and *Ae. albopictus* can live at temperatures between -6 and 31.5°C and pH ranging from 4.2 to 9.8. According to Clark et al. (2004), the pH of water can affect the success of mosquito larvae breeding, and mosquito larvae will die at a pH of less than three and more than 12, salinity of less than 0.01 to 6.3 ppt (Medeiros et al. 2020), and humidity of 60-90% (Clements 1992).

This study observes four species of the genus *Culex*, i.e., *Cx. quinquefasciatus*, *Cx. vishnui*, *Cx. tritaenorynchus*, and *Cx. gelidus*. In general, *Culex* can survive at temperatures from 24°C to 29°C. However, survival will decrease at low temperatures ($\leq 12^{\circ}$ C), at high temperatures ($\geq 32^{\circ}$ C), and die at 35°C (Grech et al. 2013). The larvae prefer slightly alkaline water (pH<8), 60% to 98% humidity (Muturi et al. 2009), and the highest salinity tolerance of 6 ppt (Fakhriedzwan et al. 2011).

The Anopheles larvae found in this study was An. vagus, which lives in rice fields. The pH value in Biringkanaya, Pattalassang, Bontoramba, and Balocci is 6.7 to 7.58. According to Maretasari et al. (2019), the pH value of rice fields is 6-8. Akhiriyanti and Nugroho (2019) reported that mosquito larvae prefer a pH range of 6.8-7.2. The temperature ranged from 26.9°C to 34.2°C in the fields where An. vagus larvae were discovered. According to Novianto et al. (2021) and Kengluecha et al. (2005), larvae of An. vagus can live in water temperatures ranging from 25.4°C to 32°C. Rice fields are areas without shade so that sunlight can penetrate the water and provide a higher temperature value than areas shaded by trees. Warm water due to sunlight may be a determining factor for larval development and accelerate the growth of Anopheles sp. In addition, warm temperatures allow more microorganisms as a food source for mosquito larvae (Minakawa et al. 1999).

The density of larvae has a significant effect on their breeding sites, such as *Ae. aegypti*, *Ae. albopictus*, *Cx. quinquefasciatus*, *Cx. visnui*, and *Cx. tritaenorynchus*. The density of mosquito larvae correlates with the conditions of their breeding sites (Medeiros et al. 2020; Vanlalruia et al. 2014). According to Madzlan et al. (2016), in addition to the factors of breeding sites, the physical condition of the water influences the density of mosquito larvae. Due to limited data on larval breeding sites and number of larvae found, the correlation cannot be determined for *Cx. gelidus* and *An. vagus* in this study.

Several studies have found that the survival of this species *Culex* sp. is linked to the habitat's environmental conditions, predators, pH, rainfall, soil conditions, aquatic plants, and sunlight intensity (Klinkenberg et al. 2003; Mutero et al. 2000; Mwangangi et al. 2006). Another factor to consider is the turbidity of the water, which influences mosquitoes' breeding attractiveness. Insoluble soil particles, organic matter, microorganisms, and other materials contribute to water turbidity (Kean et al. 2015; Munga et al. 2005). Mosquitoes are a formidable disease vector because they are found worldwide and adapt to various environmental conditions (Chandrasegaran et al. 2020). Forests, mountains, plains, deserts, tropical forests, salt marshes, and tidal zones are among the habitats where the species can breed (Foster & Walker 2019). This study concludes that mosquitoes can breed in various types of neglected water containers. Therefore, public awareness of mosquito larvae ecology in the environment is essential for reducing the density and intensity of vectors as arbovirus carriers in humans.

CONCLUSION

The application of vector control methods generally focused on the environment of urban areas. The availability of mosquito larvae in remote areas could have health consequences through neglected arbovirus and parasite disease incidence. Therefore, the results of this study are expected to be input into the implementation of policies related to vector control which are not only based on the location where DHF patients are found but also consider vector findings based on larvae surveys found in remote areas.

ACKNOWLEDGEMENTS

The authors wish to thank PMDSU Scholarship from the Indonesian Ministry of Education and Culture and the Faculty of Medicine, Hasanuddin University, Makassar, Indonesia, for supporting this research.

AUTHORS DECLARATIONS

Funding Statement

Research was supported by PMDSU Scholarship from the Indonesian Ministry of Education and Culture.

Conflict of Interest

The authors declare that there's no conflict of interests.

Ethics Declarations

This research did approve by the Health Research Ethics Committee of the Hasanuddin University Medical Faculty with the attached number 558/UN4.6.4.5.31/PP36/2020

Data Availability Statement

My manuscript has no associated data

Authors' Contributions

AR, IW and ARJ conceived this research and designed experiments; HA, DM and MI participated in the design and interpretation of the data; AR, MP, AIT, A and H performed experiments and analysis; AR, IW, S and ARJ wrote the paper and participated in the revisions of it. All authors read and approved the final manuscript. All authors read and approved the final manuscript.

REFERENCES

- Akhiriyanti, V. & Handoyo, W. 2019. Determinan keberadaan jentik di wilayah pedesaan endemis demam berdarah Dengue. *Jurnal Kesehatan Masyarakat Indonesia* 14(2): 24-28.
- Augustina, I., Jabal, A.R, Permana, G.I. & Ratnasari, A. 2021. Distribution and ecology of mosquito larvae in Pahandut Sub-District Palangka Raya City. *Journal of Physics: Conference Series* 1918(5): 052018.
- Arif, M., Tauran, P., Kosasih, H., Pelupessy, N.M., Sennang, N., Mubin, R.H., Sudarmono, P., Tjitra, E., Murniati, D., Alam, A., Gasem, M.H., Aman, A.T., Lokida, D., Hadi, U., Parwati, K.T.M, Chen-Yen, L., Neal, A. & Karyana, M. 2020. Chikungunya in Indonesia: Epidemiology and diagnostic challenges. *Plos Neglected Tropical Diseases* 14(6): 1-18.
- Hamidian, S.A. 2011. Larval habitat characteristics of the genus Anopheles (Diptera: Culicidae) and a checklist of mosquitoes in Guilan Province, Northern Iran. Iran J Arthropod-Borne Disease 5(1): 37–53.
- Bashar, K., Rahman, M.S., Nodi, I.J. & Howlader, A.J. 2016. Species composition and habitat characterization of mosquito (Diptera: Culicidae) larvae in semi-urban areas of Dhaka, Bangladesh. *Pathogens and Global Health* 110(2): 48-61.
- Carvajal, T.M., Capistrano, J.R., Hashimoto, Ki., Go, K.D., Cruz, M.J., Martinez, M.B., Tiopianco, V.P., Amalin, D.M. & Watanabe, K. Detection and distribution of *Wolbachia* endobacteria in *Culex quinquefasciatus* populations (Diptera: Culicidae) from Metropolitan Manila, Philippines. *Journal Vector Borne Disease* 55: 265-270.
- Chaiphongpachara, T., Yusuk, P., Laojun, S. & Kunphichayadecha, C. 2018. Environmental factors associated with mosquito vector larvae in a malaria-endemic area in Ratchaburi Province, Thailand". *The Scientific World Journal* 2018(3):1-8.
- Chandrasegaran, K., Lahondère, C., Escobar, E.L. & Vinauger, C. 2020. Linking mosquito ecology, traits, behavior, and disease transmission. *Trends in Parasitology* 36(4): 393-403.
- Clark, M.T., Flis, B.J. & Remold, S.K. 2004. pH tolerances and regulatory abilities of freshwater and euryhaline Aedine mosquito larvae. *Journal of Experimental Biology* 207: 2297-2304.
- Clements, A.N. 1992. The Biology of Mosquito. UK: CABI Publishing.
- Drakou, K., Nikolaou, T., Vasquez, M., Petric, D., Michaelakis, A., Kapranas, A., Papatheodoulou, A. & Koliou, M. 2020. The effect of weather variables on mosquito activity: A snapshot of the main point of entry of cyprus. *International Journal of Environmental Research and Public Health* 17(4): 1403.
- Fakhriedzwan, F., Ramasamy, R. & Mohammed, Y.K., 2011. Salinity tolerances of mosquito vectors of human disease in Brunei Darussalam. *Medicine and Health* 6(1): 190.

- Ferede, G., Tiruneh M., Abate, E., Kassa, W.J. & Wondimeneh, Y. 2018. Distribution and larval breeding habitats of *Aedes* mosquito species in residential areas of northwest Ethiopia. *Epidemiology and Health* 40(e2018015): 1-7.
- Fros, J.J., Miesen, P., Vogels, C.B., Gaibani, P., Sambri, V., *Martina, B.E.*, Koenraadt, C.J., Van Rij, R.P., Vlak, J.M., Takken, W. & Pijlman G.P. 2015. Comparative Usutu and West Nile virus transmission potential by local *Culex pipiens* mosquitoes in northwestern Europe. *One Health* 1: 31-36.
- Foster, W.A. & Walker, E.D. 2019. Mosquitoes (Culicidae). In. Mulen, G.R. & Durden LA. (eds.). *Medical and Veterinary Entomology*, pp. 792. Academic Press: Elsevier Inc.
- Futami, K., Iwashita, H., Higa, Y., Lutiali, P.A.,Sonye, G.O., Mwatele, C., Njenga, S.M. & Minakawa, N. 2020. Geographical distribution of *Aedes aegypti* (Diptera: Culicidae) in Kenya and environmental factors related to their relative abundance. *Journal of Medical Entomology* 57(3): 772–779.
- Golding, N., Nunn, M.A., Medlock, J..M, Purse, B.V., Vaux, A.G.C. & Schäfer, S.M. 2012. West Nile virus vector *Culex modestus* established in Southern England. *Parasit & Vectors* 5(32): 1-5.
- Grech, M., Sartor, P., Estallo, E., Ludueña-Almeida, F. & Almirón, W. 2013. Characterisation of *Culex quinquefasciatus* (Diptera: Culicidae) larval habitats at ground level and temporal fluctuations of larval abundance in Córdoba, Argentina. *Memórias do Instituto Oswaldo Cruz* 108(6): 772-777.
- Guo, X.X., Li, C.X., Zhang, Y.M., Xing, D., Yan-De, D., Heng-Duan, Z., Cheng-Feng, Q. & Zhao, T.Y 2016. Vector competence of *Aedes albopictus* and *Aedes aegypti* (Diptera: Culicidae) for the DEN2-FJ10 and DEN2-FJ11 strains of the dengue 2 virus in Fujian, China. *Acta Tropica* 161: 86-90.
- Ha, T.V., Kim, W., Nguyen-Tien, T., Lindahl, J., Nguyen-Viet. H., Thi, N.Q., et al. 2021. Spatial distribution of Culex mosquito abundance and associated risk factors in Hanoi, Vietnam. *PLoS Neglected Tropical Diseases* 15(6): e0009497.
- Hamidun, S., Shafie, F.A., Ishak, A.R. & Dom, N.C. 2021. Distribution and abundance of *Aedes* mosquito breeding sites at construction site workers' hostel in Gelang Patah, Johor, Malaysia. *Serangga* 26(3): 57-68.
- Higa, Y., Yen, T.N, Kawada, H., Son, T.H., Hoa, N.T. & Takagi, M. 2010. Geographic distribution of *Aedes aegypti* and *Aedes albopictus* collected from used tires in Vietnam. *Journal of American Mosquito Control Association* 26(1): 1–9.
- Huang, Y.J.S., Victoria, B.A., Amy, C.L., Unlu, I., Alto, B.W., Cohnstaedt, L.W., Higgs, S. & Vanlandingham, D.L. 2016. *Culex* species mosquitoes and zika virus. *Vector-Borne Zoonotic Diseases* 16(10): 673-676.
- Ibáñez-Justicia, A., Alcaraz-Hernandez, J.D., Lammeren, R.V., Koenraadt, C.J.M, Bergsma, A., Delucchi, L., Rizzoli, A. & Takken, W. 2020. Habitat suitability modelling to assess

the introductions of *Aedes aegypti* (Diptera: Culicidae) in the Netherlands. *Parasites and Vector* 13(217): 1-13.

- Inunggita, R., Saraswati, L.D. & Martini. 2019. Breeding places characteristic of Anopheles mosquito in Bagelen subdistrict, Purworejo. *IOP Conf. Series: Earth and Environmental Science* 246: 012053.
- Jiero, S. & Pasaribu, A.P. 2021. Haematological profile of children with malaria in Sorong, West Papua, Indonesia. *Malaria Journal* 20(126): 1-12.
- Jones, K.E., Patel, N.G., Levy M.A., Storeygard, A. & Balk, D. 2008. Global trends in emerging infectious diseases. *Nature* 451(7181): 990-3.
- Kean, J., Rainey, S.M., McFarlane, M., Donald, C.L, Schnettler, Kohl, A. & Pondeville, E. 2015. Fighting arbovirus transmission: Natural and engineered control of vector competence in *Aedes mosquitoes. Journal of Insects* 23:6(1): 236-78.
- Kengluecha, A., Singhasivanon, P., Tiensuwan, M., Jones, J.W., Sithiprasasna, R. 2005. Water quality and breeding habitats of anopheline mosquito in northwestern Thailand. *Southeast Asian Journal of Tropical Medicine & Public Health* 36(1): 46-53.
- Kesetyaningsih, T.W., Andarini, S., Sudarto & Pramoedyo, H. 2018. Determination of environmental factors affecting dengue incidence in Sleman district, Yogyakarta, Indonesia. *African journal of infectious diseases* 12(1 Suppl): 13–25.
- Khariri. 2018. Short Communication: Diversity of mosquitoes in Central Java, Indonesia that act as new vector in various tropical diseases. *Bonorowo Wetlands* 8(2): 71-74.
- Kitvatanachai, S. Janyapoon K., Apiwathnasorn, C. & Leemingsawat, S. 2005. Distribution of medically important mosquitoes in Nava Nakorn industrial estate, Pathum Thani Province, Thailand. *Journal of Tropical Medicine and Parasitology* 28(1): 8–15.
- Klinkenberg, E., Takken, W., Huibers, F. & Toure, Y.T. 2003. The phenology of malaria mosquitoes in irrigated rice fields in Mali. *Acta Tropica* 85:71-85.
- Lam-Phua, S.G., Yeo, H., Lee, R.M.L., Chong, C.S., Png, A.B., Foo, S.Y., Liew, C., Ng., L-C., Tang, C-S., Rueda, L.M. Pecor., J.E. & Harrison, B.A. 2019. Mosquitoes (Diptera: Culicidae) of Singapore: Updated Checklist and New Records. *Journal of Medical Entomology* 56(1): 103–119.
- Lindahl, J., Chirico, J., Boqvist, S., Thu, H.T.V. & Magnusson, U. 2012. Occurrence of Japanese encephalitis virus mosquito vectors in relation to urban pig holdings. American Journal of Tropical Medicine and Hygiene 87(6): 1076-1082.
- Low, V.L., Chen, C.D., Lee, H.L., Lim, P.E., Leong, C.S. & Sofian-Azirun, M. 2012. Nationwide distribution of culex mosquitoes and associated habitat characteristics at residential areas in Malaysia. *Journal of the American Mosquito Control Association* 28(3): 160–169.

- Lubinda, J., Treviño, C.J.A., Walsh, M.R., Moore, A.J., Hanafi-Bojd, A.A., Akgun, S., Zhao, B., S. Barro, A.S., Begum, M., Jamal, H, Angulo-Molina, A. & Haque, U. 2019. Environmental suitability for *Aedes aegypti* and *Aedes albopictus* and the spatial distribution pattern of major arboviral infections in Mexico. *Journal Parasite Epidemiology Control* 12(6): e00116.
- Madzlan, F., Dom, N.C, Tiong, C.S. & Zakaria, N. 2016. Breeding characteristics of Aedes mosquitoes in dengue risk area. *The Procedia Social and Behavioral Sciences* 234: 164-172.
- Maretasari, G., Windusari., Yuanita, L., Hanum, L. & Septiawati, D. 2019. Characteristics of habitat, distribution, and diversity of *Anopheles* spp in Kemelak Bindung Langit Village, Ogan Komering Ulu Regency, South Sumatra. *Journal of Environmental Science and Sustainable Development* 2(2): 165-175.
- Medeiros, S.R., Oliveira, C.R., Camargo, A.A., Scinachi, C.A., Milani, G.M. Urbinatti, P.R., Natal, D., Ceretti-Junior, W. & Toledo Marrelli, M.T. 2020. Influence of water's physical and chemical parameters on mosquito (Diptera: Culicidae) assemblages in larval habitats in urban parks of Sao Paulo, Brazil. *Journal Acta Tropica* 205: 105394.
- Minakawa, N., Mutero, C.M., Githure, J.I., Beier, J.C. & Yan, G. 1999. Spatial distribution and habitat characterisation of anopheline mosquito larvae in Western Kenya. *The American Journal of Tropical Medicine and Hygiene* 61(6): 1010-1016.
- Ministry of Health Indonesia. 2017. *Guidelines for Dengue Fever Entomology Survey and Key* to Identification of Aedes Mosquitoes. Jakarta: Ministry of Health Indonesia.
- Ministry of Health Indonesia. 2018. *Situasi Penyakit Deman Berdarah Di Indonesia Tahun 2017*. Indonesia: Ministry of Health Indonesia.
- Munga, S., Minakawa, N., Zhou, G., Barrack, O.J., Githeko, A.K. & Yan, G. 2005. Oviposition site preference and egg hatchability of *Anopheles gambiae*: Effects of land cover types. *Journal of Medical Entomology* 42(6): 993-997.
- Mutero, C.M., Blank, H., Konradsen, F. & Hoek, W.V.D. 2000. Water management for controlling the breeding of Anopheles mosquitoes in rice irrigation schemes in Kenya. *Acta Tropica* 76: 253-263.
- Muturi, E.J., Mwangangi, J., Shilihi, J., Muriu, S., Jacob, B., Mbogo, C.M., John, G. & Novak, R. 2007. Evaluation of four sampling techniques for surveillance of *Culex quinquefasciatus* (Diptera: Culicidae) and other mosquitoes in Africa rice agroecosystems. *Journal of Medical Entomology* 44(3): 503–508.
- Muturi, E.J., Mwangangi, J.M., Jacob, B.G., Shililu, J.I., Mbogo, C.M., Githure, J.I., & Novak, R.J. 2009. Spatiotemporal dynamics of immature culicines (subfamily Culicinae) and their larval habitats in Mwea Rice Scheme, Kenya. *Parasitology Research* 104(4): 851-9.

- Myint, K.S.A., Kosasih, H., Artika, I.M., Perkasa, A. & Puspita, M. 2014. Short report: West nile virus documented in indonesia from acute febrile illness specimens. *American Journal of Tropical Medicine and Hygiene* 90(2):260–262.
- Mwangangi, J., Shililu, J., Muturi, E, Gu, W., Mbogo, C., Githure, J.I. & Novak, R.J. 2006. Dynamics of immature stages of *Anopheles arabiensis* and other mosquito species (Diptera: Culicidae) in relation to rice cropping in a rice agro-ecosystem in Kenya. *Journal of Vector Ecology* 31(2): 241-245.
- Nchoutpouen, E., Talipouo, A., Djiappi-Tchamen, B., Djamouko-Djonkam, L., Kopya, E., Ngadjeu, C.S., Doumbe-Belisse, P., Awono-Ambene, P., Kekeunou, S., Wondji, C.S. & Antonio-Nkondjio, C. 2019. Culex species diversity, susceptibility to insecticides and role as potential vector of lymphatic filariasis in the city of Yaoundé, Cameroon. *PLoS Neglected Tropical Diseases* 13(e0007229): 1-6.
- Ng, S.H., Homathevi, R. & Chua, T.H. 2016. Mosquitoes of Kudat: Species composition and their medical importance (Diptera: Culicidae). *Serangga* 21(2): 149-162.
- Novianto, D., Alya, S., Hadi, U.K. & Soviana, S. 2021. Distribution and the habitat characteristics of *Anopheles vagus* (Diptera: Culicidae) larvae at paddy fields in the vicinity of dramaga IPB university campus dramaga Bogor West Java. *Acta Veterinaria Indosiana* special issue: 137-141.
- O'Connor, C.T. & Soepanto, A. 1999. Buku Kunci Identifikasi Larva dan Nyamuk Anopheles spp di Indonesia. Direktorat Jenderal Pemberantasan Penyakit Menular dan Penyehatan Lingkungan Pemukiman. Indonesia: Kementerian Kesehatan Indonesia.
- Patriani, D., Arguni, E., Kenangalem, E., Dini, S., Sugiarto, P. Hasanuddin, A., Lampah, D.A., Douglas, N.M., Anstey, N.M., Simpson, J.A., Price, R.N. & Poespoprodjo, J.R. 2019. Early and late mortality after malaria in young children in Papua, Indonesia. *BMC Infectious Diseases* 19(922): 1-13.
- Rahma, N., Hasan, H., Ratnasari, A. & Wahid, I. 2020. The application of novel methods of animal barrier screen and kelambu trap for mosquitoe's surveillance in South and West Sulawesi, Indonesia. *Biodiversitas* 2(10): 4787-4794.
- Ratnasari, A., Jabal, A.R., Rahma, N., Rahmi, S.N., Karmila, M. & Wahid, I. 2020. The ecology of *Aedes aegypti* and *Aedes albopictus* larvae habitat in coastal areas of South Sulawesi, Indonesia. *Biodiversitas* 21 (10): 4648-4654.
- Ratnasari, A., Jabal, A.R., Syahribulan, Idris, I. & Rahma, N. 2021. Salinity tolerance of larvae *Aedes aegypti* inland and coastal habitats in Pasangkayu, West Sulawesi, Indonesia. *Biodiversitas* 22(3): 1203-1210.
- Reuben, R., Thenmozhi, V., Samuel, P.P., Gajanana, A. & Mani, T.R. 1992. Mosquito blood feeding patterns as a factor in the epidemiology of JE in Southern India. *American Journal of Tropical Medicine and Hygine* 46(6): 654–663.
- Rezza, G., Chenb, R. & Weaverb, S.C. 2017. O'nyong-nyong fever: A neglected mosquitoborne viral disease. *Pathogens and Global Health* 11(6): 271–275.

ISSN 1394-5130

- Rueda, L.M. 2004. Pictorial keys for the identifications of mosquitoes (Diptera: Culicidae) associated with dengue virus transmission. *Zootaxa* 589(1): 1-60.
- Robert, V., Goff, G.L., Ariey, F. & Duchemin, J.B. 2002. A possible alternative method for collecting mosquito larvae in rice fields. *Malaria Journal* 1(4): 1-4.
- Satoto, T.B.T., Umniyati, S.R., Astuti, F.D., Wijayanti, N., Gavotte, L. Devaux, C. & Roger, F. 2014. Assessment of vertical dengue virus transmission in *Aedes aegypti* and serotype prevalence in Bantul, Indonesia. *Asian Pacific Journal Tropical Disease* 4(2): 563-568.
- Shaman, J., Day, J.F. & Komar, N. 2010. Hydrologic conditions describe West Nile virus risk in Colorado. *Journal Environmental Research and Public Health* 7(2): 494–508.
- Widarso, H., Purba, W., Suroso, T., Ganefa, S., Hutabarat, T, Cicilia, W. & Burni, E. 2002. Current status on *Japanese encephalitis* in Indonesia. Annual Meeting of the Regional Working Group on Immunization in Bangkok.
- Vanlalruia, K., Senthilkumar, N. & Gurusubramanian, G. 2014. Diversity and abundance of mosquito species in relation to their larval habitats in Mizoram, North Eastern Himalayan region. Acta Tropica 137: 1-18.
- Wharton, R.H., Eyles, D.E., Warren, M., Moorhouse, D.E. & Sandosham, A.A. 1963. Investigations leading to the identification of members of the *Anopheles umbrosus* group as the probable vectors of mouse deer malaria. *Bulletin of World Health* 29(30): 357–374.